'Silencing of host *ParPMC* genes confers resistance to *Plum Pox* Virus (PPV) in apricot (*Prunus armeniaca* L.)'

instituto valenciano de investigaciones agrarias

M.L. BADENES¹, C. ROMERO², J. BLANCA³, E. ZURIAGA^{1*}

¹Instituto Valenciano de Investigaciones Agrarias (IVIA), Valencia, Spain ²Instituto de Biología Molecular y Celular de Plantas (IBMCP-CSIC), Valencia, Spain

³Instituto de Conservación y Mejora de la Agrodiversidad Valenciana (COMAV), Valencia, Spain

*garcia_zur@gva.es

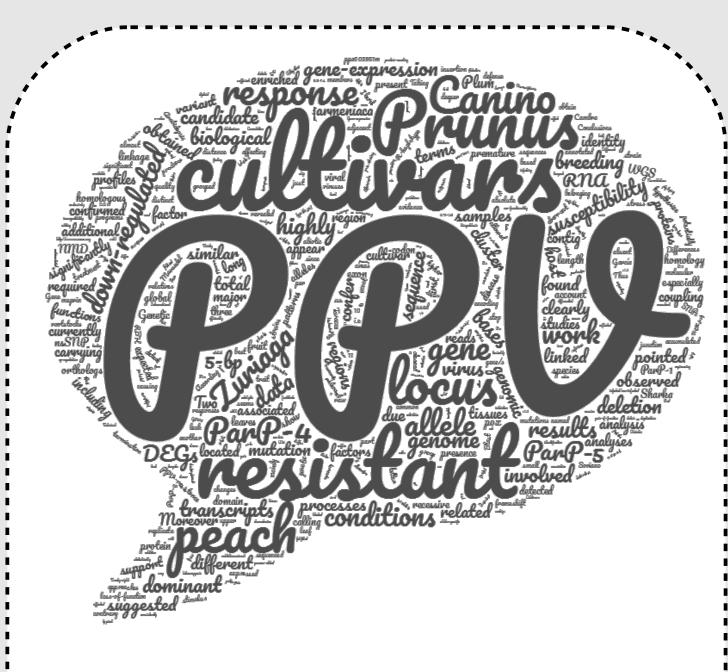
Background:

Plum pox virus (PPV), causing Sharka disease, is one of the main limiting factors for *Prunus* production worldwide. In apricot (*Prunus armeniaca* L.) the major PPV resistance locus (*PPVres*), comprising ~ 196 kb, has been mapped to the upper part of linkage group 1. Within the *PPVres*, 68 genomic variants linked in coupling to PPV resistance were identified within 23 predicted transcripts according to peach genome annotation. Taking into account the predicted functions inferred from sequence homology, some members of a cluster of meprin and TRAF-C homology domain (MATHd)-containing genes were pointed as PPV resistance candidate genes.

Results:

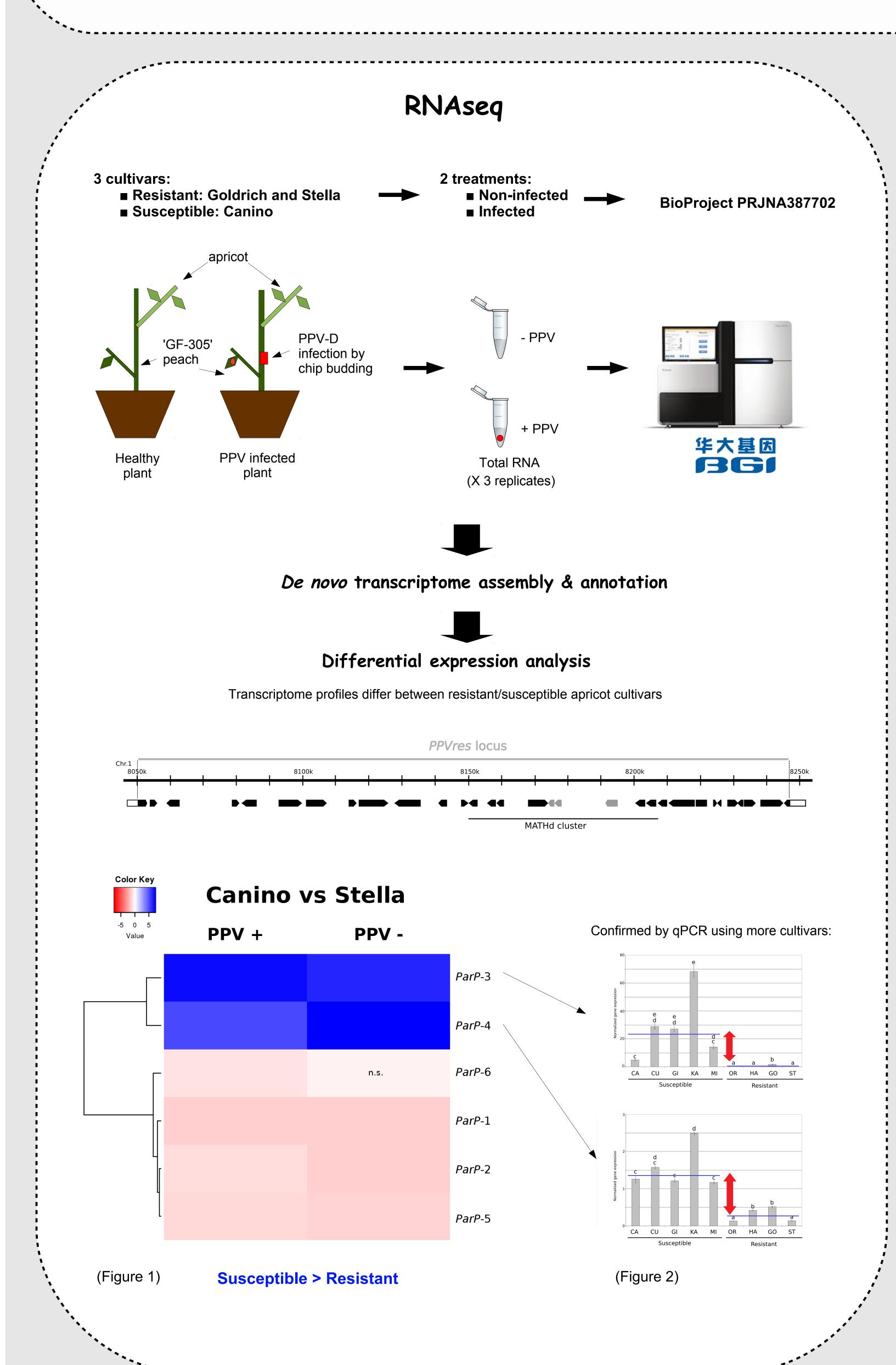
Here, we have characterized the global apricot transcriptome response to PPV-D infection. None of the *PPVres* locus genes was differentially expressed between infected/non-infected tissues, suggesting that PPV-D presence does not modulate their expression. However, six *PPVres* locus genes (*ParP-1* to *ParP-6*) appeared differentially expressed in resistant/susceptible cultivars (Figure 1). Two of them (*ParP-3* and *ParP-4*), that encode MATHd proteins, appear clearly down-regulated in resistant cultivars, as confirmed by qRT-PCR (Figure 2). Concurrently, variant calling was performed using whole-genome sequencing data of 24 apricot cultivars (10 PPV-resistant and 14 PPV-susceptible) and 2 wild relatives (PPV-susceptible) (Figure 3). *ParP-3* and *ParP-4*, named as *Prunus armeniaca PPVres MATHd-containing genes (<i>ParPMC1* and *ParPMC2*, respectively), are the only 2 genes having allelic variants linked in coupling to PPV resistance. *ParPMC1* has 1 nsSNP, while *ParPMC2* has 15 variants, including a 5-bp deletion within the second exon that produces a frameshift mutation. *ParPMC1* and *ParPMC2* are highly homologous (87.5% identity) and adjacent in the MATHd genes cluster suggesting they are paralogs originated from a tandem duplication (Figure 4). Cultivars carrying the *ParPMC2* resistant (mutated) allele show lack of expression in both *ParPMC2* and especially *ParPMC1*.

Conclusions: Accordingly, we hypothesize that *ParPMC2* is a pseudogene that mediates down-regulation of its functional paralog *ParPMC1* by silencing. As a whole, results strongly support *ParPMC1* and/or *ParPMC2* as host susceptibility genes required for PPV infection which silencing may confer PPV resistance trait. This finding may facilitate resistance breeding by marker-assisted selection and pave the way for gene edition approaches in *Prunus*.



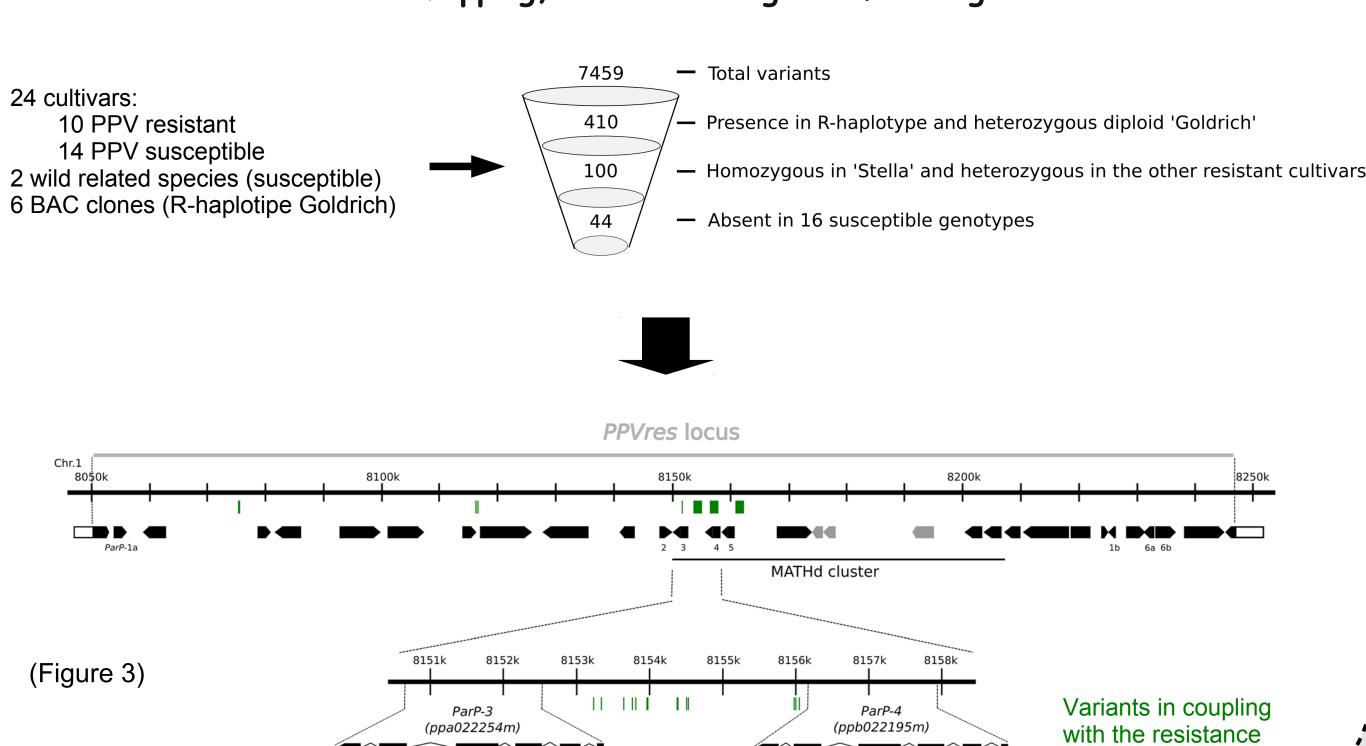
This work has been published as:

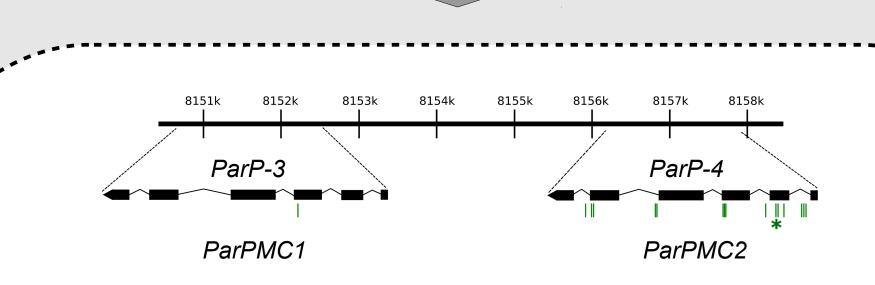
Zuriaga, et al. (2018). Resistance to *Plum Pox Virus* (PPV) in apricot (*Prunus armeniaca* L.) is associated with down-regulation of two *MATHd* genes. BMC Plant Biology 18:25.



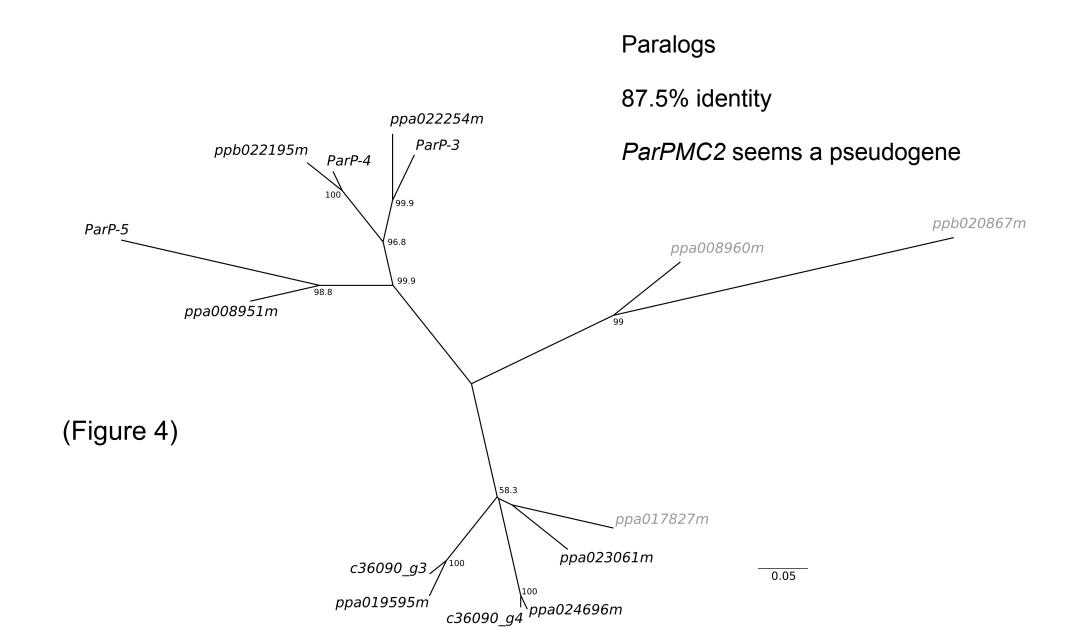
Whole-genome sequencing

WGS mapping, variant calling and filtering





<u>Prunus armeniaca PPVres MATHd-containing genes (ParPMC)</u>
Host susceptibility genes required for PPV infection



Down-regulation of *ParPMC1* and *ParPMC2* gene expression is the differential factor between PPV resistant and susceptible apricot cultivars regarding the *PPVres* locus. **How?**

1) Mutations in the promoter regions of the PPV resistant alleles may be directly affecting their expression, but half-gene dosage can not account for the observed differences between resistant and susceptible cultivars.

2) ParPMC1 and ParPMC2 gene expression may also be prevented by **gene-silencing**. How? Non-mediated decay? Pseudogene-silencing his protein-coding cousin?

More analyses are in progress to elucidate this mechanism

Acknowledgments

We thank University of Málaga (Spain) for data processing through the Picasso supercomputer. We acknowledge Dr. Rios and Alba Lloret for advice on qPCR, Dr. Dardick, Dr. Zhebentyayeva and Dr. Abbot for providing some WGS data and Dr. Cañizares for his helpful comments on the manuscript.

This research was supported by the Spanish Ministry of Economy, Industry and Competitiveness (Research Project RTA2013–00026-C03–01).