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1Application of a new wax containing ethanol as a method to remove persimmon

2astringency during cold storage

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25experiments; AFN carried out the experiment, analysed the data and wrote the MS.

26

27

28 **Research highlights**

29 • Wax containing ethanol is effective to remove packaged persimmon astringency
30 during cold storage

31 • CO₂ deastringency treatment may cause quality loss and flesh browning after
32 prolonged cold storage

33 • New patented method can replace CO₂ deastringency treatment when prolonged cold
34 storage is needed

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43Abstract

44Nowadays the treatment based on applying high CO₂ concentrations to fruit is the main
45method used in astringent persimmon prior to being commercialized, but it can cause quality
46problems for fruit during cold storage. The aim of this study was to evaluate the effectiveness
47of a recently patented astringency removal method based on applying a new wax whose
48formulation includes ethanol before commercial packaging. During two seasons, three
49treatments were evaluated in cv. Rojo Brillante and Triumph: 1) CO₂- standard treatment; 2)
50waxed and packed in plastic film according to the patented method; 3) packed in plastic film
51without any treatment. During a third season, the new method's effectiveness in removing
52astringency was evaluated under industrial conditions. After treatments fruit was stored at
530°C for 15, 21 and 30 days before being transferred at 20°C to simulate a 5-days shelf-life.
54All the fruit treated with the new wax completely lost astringency after 30 days at 0°C, and
55commercial firmness was maintained. At the end of the storage, fruit quality was
56substantially higher in fruit submitted to the new treatment. CO₂-treated fruit, manifested
57internal browning after 30 storage days and shelf-life, while this disorder was not detected in
58waxed fruit.

59**Keywords: deastringency, cold storage, wax, ethanol, 'Rojo Brillante', 'Triumph'**

62Introduction

63Persimmon cultivars are divided into two categories, astringent and non-astringent,
64depending on their astringency level at harvest. Astringent persimmons are not edible when
65harvested due to the presence of high soluble tannin concentrations, which produce a dry
66puckering sensation in the mouth (Besada and Salvador 2018). ‘Triumph’ and ‘Rojo
67Brillante’ are the main cultivars from the Mediterranean region, both of which are astringent
68at harvest.

69The most habitual treatment used to remove astringency in persimmon is based on exposing
70fruit to 95-98% CO₂ at 20 °C and 90% RH for 24 h (Besada and Salvador 2018). The
71effectiveness of this method lies in the fact that it triggers anaerobic respiration in fruit, which
72gives rise to the accumulation of acetaldehyde. The reaction between this acetaldehyde and
73the soluble tannins responsible for astringency leads tannins to become insoluble, hence they
74lose their astringent character (Arnal and Del Río 2003; Salvador et al. 2007). Although the
75effectiveness of this deastringency method has been widely studied, a high CO₂ concentration
76affects the parenchyma structure, causing cell membrane degradation, which can lead to loss
77of flesh firmness after treatment. Thus, in the case the fruit has to be cold stored, it is
78recommended to remove the astringency after conservation to preserve the fruit firmness.
79Nevertheless, it is noteworthy that in some marketing scenarios, such as shipments to
80overseas countries, CO₂ treatment is necessarily applied before using refrigerated transport.
81Under these conditions, the risk of fruit quality loss during cold storage is high. Indeed, the
82fruit softening and internal browning that appears during prolonged refrigerated transport is
83one of the most important problems that is limiting the market to overseas countries.

84In this context, interest has been shown in finding alternative deastringency methods that can
85be applied in cold storage management scenarios to avoid quality losses associated with the
86currently employed CO₂ treatment. To respond to this demand, the present work takes as a

87starting point two postharvest technologies used for different purposes that have never been
88combined. On the one hand, it is known that exposing fruit to ethanol has an effect of
89removing astringency. Ethanol vapor was one of the first treatments to be assayed to remove
90astringency in the persimmon fruit industry (Taira et al. 1989). Although this treatment still
91continues to be used in countries like Brazil (Tessmer et al. 2018), it is being replaced with
92CO₂ treatment as the latter requires less time to remove astringency and fruit firmness is
93better preserved.

94On the other hand, edible waxes are used to carry some ingredients like antioxidants,
95antifungal and antimicrobial agents for different purposes in different fruits (Nair et al. 2018;
96Rojas-Graü et al. 2007). In persimmon no information on this regard exists, only some studies
97have reported the use of waxes to reduce softening and weight loss (Blume et al. 2008;
98Carvalho da Silva et al. 2011).

99In this context, the hypothesis based on producing an edible wax as a carrier for ethanol was
100the starting point to reach research collaboration between the Instituto Valenciano de
101Investigaciones Agrarias (IVIA) and the Fomesa Fruitech S.L.U. Company. The final
102objective was to find a new deastringency method based on ethanol that can be easily applied
103and that does not have negative effects on the fruit quality during low-temperature storage
104and subsequent shelf-life. The studies carried out allowed to develop an ethanolic wax that
105could remove fruit astringency during cold storage when combined with fruit packaging, and
106this process has been recently patented (P-101459).

107The objective of this work was to evaluate the effectiveness of this new method to remove
108persimmon astringency during cold storage, and to determine its effect on other important
109physico-chemical quality parameters as color, firmness, total soluble solids, chilling injury
110and browning incidence. This study was part of the studies submitted for approval of the
111patent.

112

113Materials and Methods

114 **Fruit source and treatments**

115 This study was conducted in three consecutive seasons from 2017 to 2019. In the first and
116 second season the treatments were tested under lab conditions while in the third season the
117 assays were carried out under industrial conditions.

118 First season (S1). ‘Rojo Brillante’ persimmons were harvested in November in a commercial
119 maturity stage from an orchard in Valencia (Spain) before being immediately transported to
120 the IVIA’s Postharvest Laboratory Technology Center. Fruit were carefully selected for their
121 uniform size and external color. Then they were separated into homogenous lots of 12 fruits,
122 and were placed on plastic alveoli trays in a cardboard fruit box. Three of these lots were
123 used to evaluate the physico-chemical quality of fruit at harvest.

124 The remaining lots of fruit were subjected to the following three treatments (12 lots per
125 treatment):

126 1) fruit submitted to the traditional CO₂ treatment with 95% CO₂ at 20°C for 24h (CO₂);

127 2) fruit waxed and packed in plastic film according to the patented method (Wax+Film);

128 3) fruit packed in plastic film with no wax application, used as the control (Film).

129 After applying treatments, all the fruit were stored at 0°C with 85%-90% RH for up to 30
130 days. Periodically after 15, 21 and 30 days, three lots of each treatment were removed from
131 cold storage for analysis. In addition, after 30 days three other lots of fruit were transferred to
132 20 °C for 5 days to simulate the shelf-life period, after which they were evaluated.

133 Second season (S2). During this season, experiments were carried out in the same way as
134 during the first season, but in this case they were performed with persimmons ‘Rojo
135 Brillante’ and ‘Triumph’. The fruit of both cultivars were harvested in the commercial
136 maturity stage in December from two orchards in Valencia. During this season, a 5-day shelf-
137 life at 20°C was simulated after 15, 21 and 30 days of cold storage periods.

138 Third season (S3). During this season, the effectiveness of the new method to remove
139 astringency was evaluated under industrial conditions with ‘Rojo Brillante’. To this end, after

140harvest fruit were transported to a commercial packing house (Natural Hand S.L.) where they
141were carefully selected for their uniform size and external color. Then fruit were separated
142into homogenous lots of 16 fruits and were placed on plastic alveoli trays in a commercial
143cardboard fruit box. Four treatments were assayed: 1) fruit submitted to the traditional CO₂
144treatment with 95% CO₂ at 20 °C for 24 h (CO₂) in industrial chambers; 2) fruit waxed and
145packed in plastic bags according to the patented method (Wax+Film); 3) fruit packed in
146plastic films with no wax application, used as the control (Film); 4) fruit waxed and boxes
147stacked on a wooden pallet, strapped with macroperforated stretch film by a commercial
148automatic strapping machine (Wax+Pallet). Here the objective was to generate a kind of film
149package by strapping pallets, whereby a cardboard sheet was placed on top of the pallet
150(Wax+ Pallet) and it was film-strapped until almost the whole pallet surface was covered.
151After applying treatments, boxes were placed in the cold chambers at the Natural Hand
152Company at 0-1 °C with 85%-90% RH for up to 30 days. After 30 days, three lots of each
153treatment were evaluated. Three other lots of fruit were transferred to 20 °C for 5 days to
154simulate the shelf-life period, after which they were evaluated.

155In all cases after harvest the fruit were treated with 1-MCP under commercial conditions (500
156nL/L of 1-MCP for 24 h at room T). This treatment is commercially applied to persimmons
157before they are submitted to low temperature in order to delay chilling injury symptoms
158(Salvador et al. 2004).

159CO₂ treatment was carried out in closed containers (467 L), which contained 95% CO₂ for 24
160h at 20 °C and 90% RH. These conditions were established by passing a stream of air
161containing 95% CO₂ through containers.

162In the first and second season, in the lab, wax was applied uniformly to the fruit surface with
163a sprayer at a rate of 12.5 mL wax per kg of fruit. Wax and plastic film (Xtend[®], StePac L.A.
164Ltd) were provided by the Fomesa Fruitech S.L.U. The application system for the wax in the
165industry (third season) was composed of two main parts, the pressurizer pump and the

166spraying gun. The main objective of this application system was to create a fine nebulization
167of the wax, creating a thin coating of the product on the surface of the fruit. The pressurizer
168pump is in charge of using pressured air from a compressor to pressurize the wax through an
1698 mm pipe to the spraying guns. Once the wax is pressured, the spraying guns are able to
170make a fine nebulization of the wax on the surface of the fruit. Regarding the spraying gun,
171we used an 80° R nozzle, which allows us to make a diffusion of the wax at an 80° opening
172angle and a flow rate of 2.5 gallons per hour.

173At harvest, and after different cold storage periods as well as the shelf-life, the determined
174physico-chemical and sensory parameters were as follows: external color, firmness, soluble
175tannins content (ST), sensory astringency and off-flavors, acetaldehyde (AcH) and ethanol
176(EtOH) in juice. After cold storage, the CO₂ and EtOH concentrations inside plastic films
177were measured.

178For a better understanding, the treatments and conditions tested in each season are shown in
179the table 1.

180

181**Fruit quality assessments**

182Determinations were made with 12 fruits, three replicates per treatment. After the low-
183temperature storage periods and the subsequent shelf-life, the following were evaluated:
184external color, flesh firmness, soluble tannin contents (ST), sensory astringency and off-
185flavors, internal and external disorders, AcH and EtOH in juice, EtOH and CO₂ in plastic
186bags.

187Skin color was determined by a Minolta colorimeter (Model CR-300; Minolta, Ramsey, NY).
188Hunter parameters “L,” “a,” and “b,” were measured and the results were expressed as Color
189Index = (1000a)/(Lb) (Salvador et al. 2007). Two measurements were taken on the opposite
190equatorial area of each fruit.

191Flesh firmness was evaluated by a texturometer (model 4301, Instron Corp., Canton, Mass.,
192USA) using an 8-mm plunger. The crosshead speed during firmness testing was set at 10 mm/
193min. The results were expressed as load in Newton (N) to break flesh at two equidistant
194locations in the equatorial region of each fruit after epicarp removal.

195Immediately after taking the firmness measurements, fruits were cut into half and any internal
196disorder was evaluated. The incidence of external and internal disorders was evaluated
197visually on all the fruit in each lot according to Khademi et al. (2013). Skin browning severity
198was evaluated on four scales as: 0, no browning; 1, slight browning; 2, moderate browning; 3,
199severe browning. Similarly, flesh browning was rated on a 4-point scale according to the
200browning intensity detected in the central fruit part as: 0, no browning; 1, slight browning
201(less than 30% of the fruit flesh surface was brown); 2, moderate browning (more than 30%
202and less than 60% of the fruit flesh surface was brown); 3, severe browning (more than 60%
203of the fruit flesh surface was brown). The browning index (BI) was calculated as BI=
204 $\sum[(\text{browning severity}) \times (\text{no. of fruit at each browning severity})] / 3 \times \text{total no. of fruit}$.

205

206After disorders evaluation, six fruits per lot were cut to obtain four quarters. Two opposite
207quarters were frozen at -21°C until the ST analyses. ST content was evaluated by the Folin-
208Denis method described by Arnal and Del Río (2004). The results were expressed as a
209percentage or g/100g FW. One of the remaining quarters was used in the sensory evaluation
210and the other was employed to obtain juice for the AcH and EtOH determinations.

211The sensory evaluation was performed by 8 trained panelists who were asked to evaluate
212astringency levels and the presence of off-flavors. Sensory sessions were carried out in a
213specifically adapted room, where panelists were seated in individual evaluation booths. Fruits
214were peeled and sliced (longitudinal slices, 1.5 cm wide), and three slices from three different
215fruits were presented per sample to compensate for variability. Water was provided to cleanse
216palates between samples. Samples were served in random order to the panelists in 50-mL
217stainless steel soufflé cups, identified by a unique three-digit number. Each fruit was tasted

218by at least three panelists. A 5-point scale was used for the sensory astringency evaluation: 1
219= no astringency; 2 = residual astringency; 3 = slight astringency; 4= moderate astringency; 5
220= astringency. A value below 1.5 guarantees the non-astringency of the fruit, and therefore is
221considered commercially acceptable (Besada et al., 2016; Munera et al., 2017). Off-flavors
222were evaluated as Presence/Absence.

223In order to determine the AcH and EtOH concentrations, one quarter of the six fruits per lot
224was placed in an electric juice extractor (model 753, Moulinex, Spain) and the obtained juice
225was filtered through cheesecloth. AcH and EtOH production was measured with three
226samples per replicate of juice samples, obtained as previously mentioned and analyzed by
227headspace gas chromatography. Five milliliters of the filtered juice were transferred to 10-mL
228vials with crimp-top caps, sealed with TFE/silicone septa, and frozen (-21) until analyzed.
229For the analysis, samples were put in a water bath at 20 °C for 1 h, followed by heating at 60
230°C for 10 min. A 1-mL headspace sample was withdrawn from the vials and injected into the
231gas chromatograph (model 2000, Perkin-Elmer, Norwalk, Conn., USA), equipped with a
232flame ionization detector (FID) and a 0.32 cm × 1.2 m Poropak QS 80/100 column. The
233injector was set at 175 °C. EtOH and AcH were identified by comparing the retention times
234with those of a standard solution. The results were expressed as mg/100 mL.

235Prior to opening the plastic bags, the EtOH and CO₂ concentrations were determined. For this
236purpose, a BD Plasticpak syringe was placed inside the plastic bag and 1 mL of the generated
237atmosphere was extracted carefully so as not to damage fruit. This volume was injected into
238the gas chromatograph (model 2000, Perkin-Elmer, Norwalk, Conn., USA) and the same
239previously explained pattern was followed. For EtOH, the results were shown as µL/L and as
240% of gas in the atmosphere for CO₂.

241Statistical Analysis

242Data were subjected to an analysis of variance, based on two factors (treatment × storage
243duration). Multiple comparisons between means were determined by the least significant

244difference test ($P \leq 0.05$) using the Statgraphics Plus 5.1 software application (Manugistics
245Inc., Rockville, MD, USA).

246

247**Results and Discussion**

248**Soluble tannins (ST) and sensorial astringency evaluation**

249

250At harvest, the ST content in ‘Rojo Brillante’ was close to 0.65-0.70% during all three
251studied seasons (Figure 1a, 1b, 1d). The ST values for ‘Triumph’ were slightly higher at
252harvest time, 0.9% (second season) (Figure 1c). These values were in the same range found
253by most previous studies conducted on both cultivars, which have been related to high
254astringency levels in fruit (Besada et al. 2014; Novillo et al. 2015; Salvador et al. 2007).

255As expected, the ST content dropped significantly after being treated with a high CO₂
256concentration (CO₂ treatment). In all cases after 1 day of the treatment (DA), the ST
257concentration lowered to values between 0.01% and 0.03%. The panelists evaluated fruit
258astringency with scores of 1 (non-astringent fruit) (Figure 1e, 1f, 1g, 1h).

259In the fruit packed in plastic film with no added wax (Film), ST slightly dropped, over all
260after 30 cold storage days and the shelf-life that followed the different cold storage periods.
261Despite of this decrease, in all cases, the ST values remained very high, which resulted in
262fruit having high levels of sensory astringency (astringency scores ranged between 3.9 and
2634.8).

264However, the ST in the waxed fruit (Wax+Film) gradually lowered during the studied storage
265period. In the first and second seasons, after 15 days at low temperature, ST drastically
266decreased, with values close to 0.15%-0.18% in both cultivars. The sensory evaluation
267revealed that this fruit still exhibited a high astringency level (scores of 4.2-4.5). In all cases
268from 15 days onward, the ST content continued to lower and reached values below 0.03%
269after 30 days, when the fruit’s astringency scores were lower than 1.5, considered to be

270commercial. Similar results were obtained during the third season, when assays were carried
271out with ‘Rojo Brillante’ under industrial conditions. After 30 cold storage days, the
272wax+film fruit and wax+pallet fruit obtained ST values close to 0.03%. In both cases, fruit
273was evaluated as non-astringent.

274One point worth mentioning is that in the second season, in which the shelf-life conditions
275were simulated after all the cold storage periods, just after 15 and 21 days at low temperature
276the wax+Film fruit values were 0.01% and 0.02% in ‘Rojo Brillante’ and ‘Triumph’,
277respectively, and the panelists did not detect astringency.

278In persimmon fruit the threshold of ST leading to astringency perception depends on the
279cultivar. ST above 0.1% has been found to confer an astringent taste, and these fruit are not
280edible in persimmons ‘Kaki Tipo’, ‘Lycopersicom’ and ‘Thiene’ (Vidrih et al. 1994). In the
281present study, and irrespectively of the treatment to which fruit was submitted, the panelists’
282evaluation of fruit not being astringent corresponded to the fruit that had an ST content below
2830.03%. This value agrees with those previously reported for both ‘Rojo Brillante’ and
284‘Triumph’ (Besada et al. 2010; 2014; Salvador et al. 2007).

285

286Acetaldehyde (AcH) and ethanol (EtOH)

287Most methods performed to remove persimmon astringency are based on maintaining fruit
288under anaerobic conditions or exposing them to products that induce anaerobic respiration.
289Under these conditions, soluble tannins, responsible for astringency, are polymerized by AcH
290that accumulates in flesh, hence the deastringency process rate has been positively related to
291the level of AcH that accumulates in fruit flesh (Pesis et al. 1988; Taira et al. 1989). In ‘Rojo
292Brillante’, the accumulated AcH concentration after different deastringency treatments has
293also been closely associated with a drop into the decrease of ST (Besada et al. 2010).
294Nevertheless, the AcH level required to insolubilize tannins in fruit flesh to reach
295undetectable astringency values has not yet been established.

296In the present study, both cultivars exhibited very low AcH concentrations at harvest,
297between 0.1 and 0.3 mg/100 mL (Figure. 2a, 2b, 2c, 2d). These values are in the range
298reported in previous studies (Arnal and Del Río 2003; 2004; Besada et al. 2014). As expected
299in the CO₂-treated fruit, AcH significantly increased 1 day after treatment (DA), with values
300of around 4.5 mg/100 mL in both cultivars in the first and second seasons. For the third
301season, the values reached after CO₂ treatment were slightly lower, 3.5 mg AcH/100 mL. For
302all three seasons, AcH levels linked to CO₂-treated fruit showed no relevant changes
303throughout the study period.

304The fruit packed in plastic film with no added wax (Film fruit) had similar values to those at
305harvest throughout cold storage. Although the AcH concentration slightly increased after the
306shelf-life periods, the maximum values did not exceed 0.7 mg/100 mL.

307In the Wax+Film fruit, AcH gradually increased throughout cold storage. So, in the first and
308second season the AcH content obtained values close to 0.8 mg/100 mL after 15 days at low
309temperature, which coincided with a significant drop in ST. From this time point, the AcH
310concentration continued to rise concomitantly with a drop in ST. After 30 cold storage days,
311when fruit was evaluated as non-astringent, the AcH content came close to 2.5 mg/100 mL
312and 3.5 mg/100 mL for 'Rojo Brillante 'and 'Triumph', respectively. It should be noted that
313similar AcH values were found during the second season (Figure 2b, 2c) in the fruit evaluated
314as being non-astringent after the shelf-life that followed the 15 and 21 cold storage days.
315Under industrial conditions (third season), loss of astringency after 30 cold storage days in
316the Wax+Film fruit was associated with an AcH concentration of 3.5 mg/100 mL.
317Nevertheless, in the palletized fruit (Wax+Pallet), this value was lower, 2 mg/100 mL (Figure
3182d).

319According to these results, an AcH concentration in fruit flesh within the 2 and 3.5 mg/100
320mL range was necessary to achieve a low enough ST content ($\leq 0.03\%$) to not induce
321perceived astringency. This AcH range is consistent with previous studies, in which similar
322AcH values have been measured in non-astringent fruit after CO₂ treatment (Besada et al.

3232010; Novillo et al. 2013; 2015; Salvador et al. 2007). In the present study, this AcH content
324was achieved only 1 day after CO₂ treatment, whereas 30 cold storage days or 15 days plus
325the shelf-life were required to reach the same AcH concentration in the waxed fruit.

326Regarding the EtOH content values went below 6 mg/100 mL at harvest in both cultivars
327(Figure 2e, 2f, 2g, 2h). In the CO₂-treated fruit, the EtOH content significantly increased 1
328day after treatment (DA). Values of 35 mg/100 mL and 65 mg/100 mL were detected in 'Rojo
329Brillante' in the first and second season respectively. The EtOH content in 'Triumph' (Figure
3302g) was higher and came close to 78 mg/100 mL. These EtOH values remained constant
331throughout cold storage and the subsequent shelf-life periods. The film fruit accumulated
332very little EtOH during storage. Nevertheless, in the Wax+Film fruit, EtOH accumulation
333was in flesh after 15 cold storage days, and values did not relevantly change until the end of
334the study. In 'Rojo Brillante', the EtOH values were 80 mg/100 mL during the first season
335and came close to 180 mg/100 mL during the second season. In 'Triumph', the EtOH values
336came close to 200 mg/100 mL. In the third season, differences in EtOH were found between
337the Wax+Film and palletized fruit (Wax+Pallet). The Wax+Film fruit showed 150 mg EtOH/
338100 ml after 30 storage days while the EtOH values of the palletized-fruit were significantly
339lower (close to 50 mg/100 mL).

340In this study, the applied wax formulation contained ethanol. Therefore, the AcH required to
341insolubilize tannins would be produced mainly by alcohol dehydrogenase directly from the
342EtOH that accumulated in flesh, as reported by other authors after studying the effectiveness
343of destringency treatments by EtOH vapor (Taira et al. 1989; Yamada et al. 2002).

344According to the EtOH measures taken inside plastic bags, an increment 1 day after cold
345storage to values of 0.5 µL/L and 0.7 µL/L was observed in the Wax+film 'Triumph' fruit
346and the 'Rojo Brillante' fruit, respectively. After 15 days however, this concentration
347dropped to those values found in plastic films with the unwaxed fruit (values close to 0.04)
348(Table 2). The low EtOH concentration that accumulated in plastic films allowed us to
349assume that the gradual increase in AcH in flesh during cold storage would be promoted by

350EtOH diffused through skin from wax into the parenchymal tissue, instead of from ethanol
351vapors. Yamasaki et al. (2017) also reported astringency removal ‘on tree’ using ethanol
352stickers to release EtOH to the fruit surface.

353It is also noteworthy that like other fruits, EtOH accumulation may play an important role in
354off-flavors developing in persimmon (Pesis 2005). In line with this, Ben-Arie et al. (1991)
355studied the effect of modified atmospheres during persimmon storage and reported how
356EtOH that accumulated to levels exceeding 75 mg/100 mL was related to deteriorated taste.
357Nevertheless, in other studies, values of 100 mg EtOH/100 mL in ‘Rojo Brillante’ or 200 mg/
358100 mL in ‘Triumph’ had no negative effect on fruit taste (Besada et al. 2014). The maximum
359EtOH values herein detected came close to 200 mg/100 mL and were, in any case, associated
360by the panelists with a negative change in fruit flavor (data not shown).

361

362**External color and flesh firmness**

363The two studied cultivars were harvested in all three seasons with homogeneous coloration
364and color index values between +8 and +11, but no relevant changes in external coloration
365were detected during the cold storage period (data not shown). Nevertheless, when fruit were
366transferred to shelf-life conditions, a significant increase took place. Thus the ‘Rojo Brillante’
367values came close to +20, and to +15 for ‘Triumph’, after the shelf-life following 30 days
368storage at 0 °C. In any case, differences among treatments were found.

369One of the most important parameters for preservation during the cold storage of persimmon
370is flesh firmness. It is stressed that most persimmon cultivars have been widely reported to
371exhibit chilling injury symptoms when stored at low temperatures, and firmness loss is the
372main chilling injury symptom. Such flesh softening can be exhibited during cold storage at 4–
37311 °C (Arnal and Río 2004). However, during storage at 0–1 °C, drastic firmness loss occurs
374mainly when fruit are transferred to shelf-life temperatures (Salvador et al. 2004; 2006). Thus
375the application of 1-MCP prior to storage has become a common treatment before submitting
376fruit to low temperatures as it reduces chilling injury symptoms in many persimmon cultivars

377(Kim and Lee 2005; Salvador et al. 2004; Tessmer et al. 2019). In the present study, all the
378fruit were submitted to commercial 1-MCP treatment after harvest to preserve flesh firmness.
379In the first and second seasons, fruit were harvested with high flesh firmness values of around
38040-45 N. In the third season, the initial firmness was slightly lower (close to 35 N). During
381cold storage, firmness slightly decreased in all cases. After transferring fruit from cold
382storage to shelf-life conditions, firmness loss became more evident, but flesh firmness
383remained crispy with values above 25 N during the first and second seasons. It was only
384during the third season when fruit underwent more marked firmness loss, with values close to
38515 N. In any case, differences among the assayed treatments were found.

386

387**Internal and external disorders**

388Throughout the study period, no external disorders were observed. Nevertheless, flesh
389browning was exhibited only in 'Rojo Brillante' subjected to CO₂ treatment after the shelf-
390life that following the 30 cold storage days (Figure 3). The Browning Index gave values of
3910.2 and 0.3 in the first and the second season, respectively (data not shown). The wax-treated
392fruit (Wax+Film) did not show any browning (Figure 3).

393Different postharvest factors have been associated with internal persimmon browning, such
394as storage in controlled atmospheres and mechanical damage (Besada et al. 2014; Novillo et
395al. 2014), and browning symptoms differ depending on the causal factor.

396The herein manifested flesh browning was visualized in the central fruit area, similarly to that
397reported to be induced by storage in controlled atmospheres (Besada and Salvador 2018). It is
398noteworthy that this alteration is not new to us because it has often been visualized in those
399persimmons submitted to CO₂ treatment before being cold-stored for lengthy periods. Indeed,
400the causes of this disorder are currently being studied by our group.

401In other fruits, tissue browning and cell death have been linked to the AcH that accumulates
402under anaerobic conditions (Fan et al. 2005; Zhang et al. 2015). Accordingly, the browning
403manifestation herein found in the fruit subjected to CO₂ treatment could be related to AcH

404accumulating in flesh. So the CO₂ fruit achieved the maximum AcH level only 1 day after
405treatment and this high AcH concentration in flesh remained constant throughout the study
406period. Nevertheless, in the wax-treated fruit, AcH accumulation was gradual and it was only
407after 30 days when it reached similar levels to those of the CO₂-treated fruit. Thus the period
408that flesh is exposed to high AcH levels seems crucial for browning development.

409

410**Conclusion**

411The patented process based on new wax is revealed as an effective deastringency treatment
412during the cold storage of persimmons ‘Rojo Brillante’ and ‘Triumph’. Complete astringency
413removal is achieved in both varieties with no fruit quality loss after 30 storage days at 0 °C
414and after the shelf-life period that followed 15 and 21 storage days. Therefore, this method is
415an alternative to CO₂ treatment when prolonged cold storage is required, as on shipments to
416far overseas markets.

417This study corroborates that applying CO₂ deastringency treatment to persimmon implies an
418important quality loss risk associated with flesh browning manifestation after prolonged cold
419storage.

420

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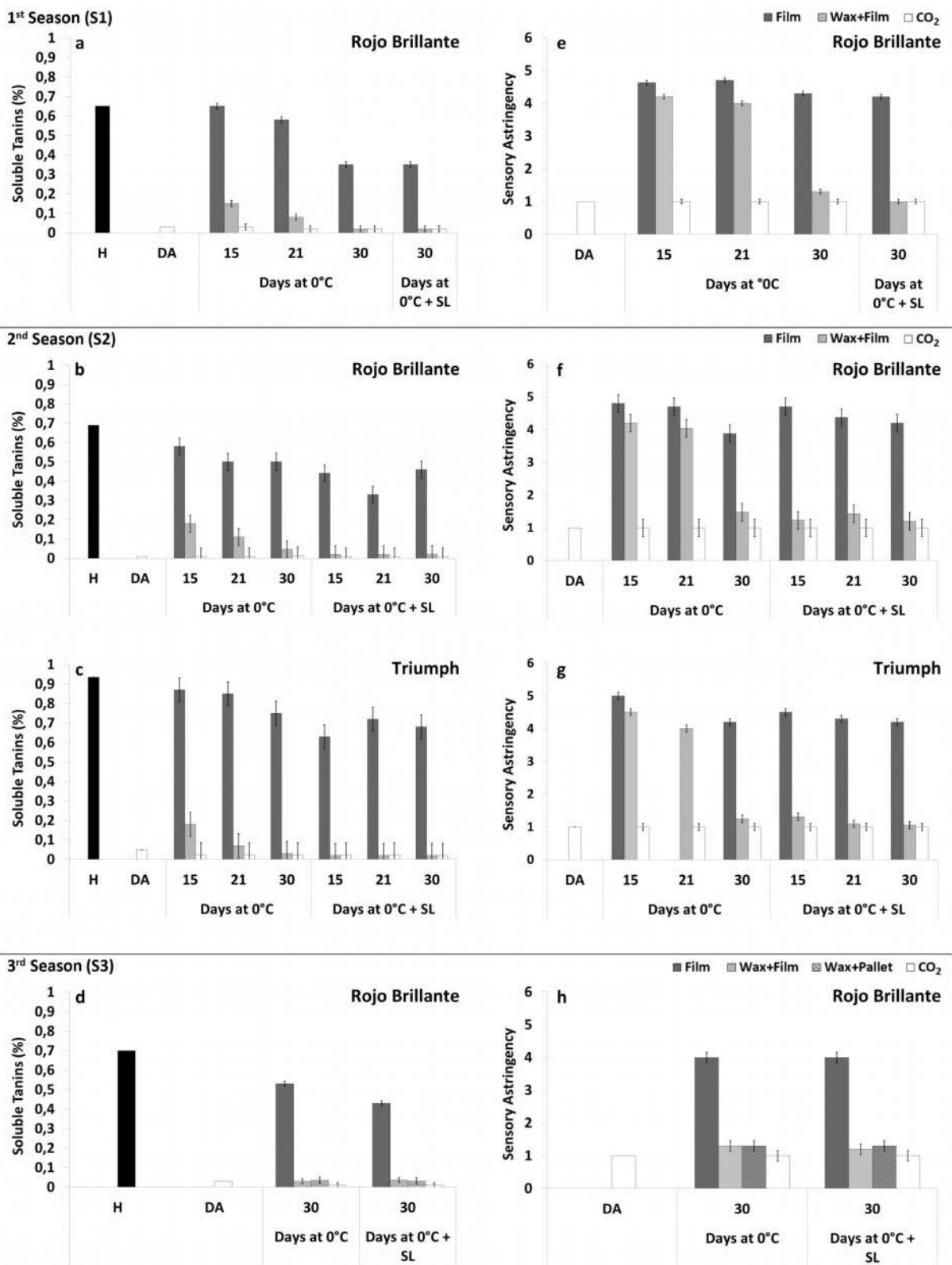
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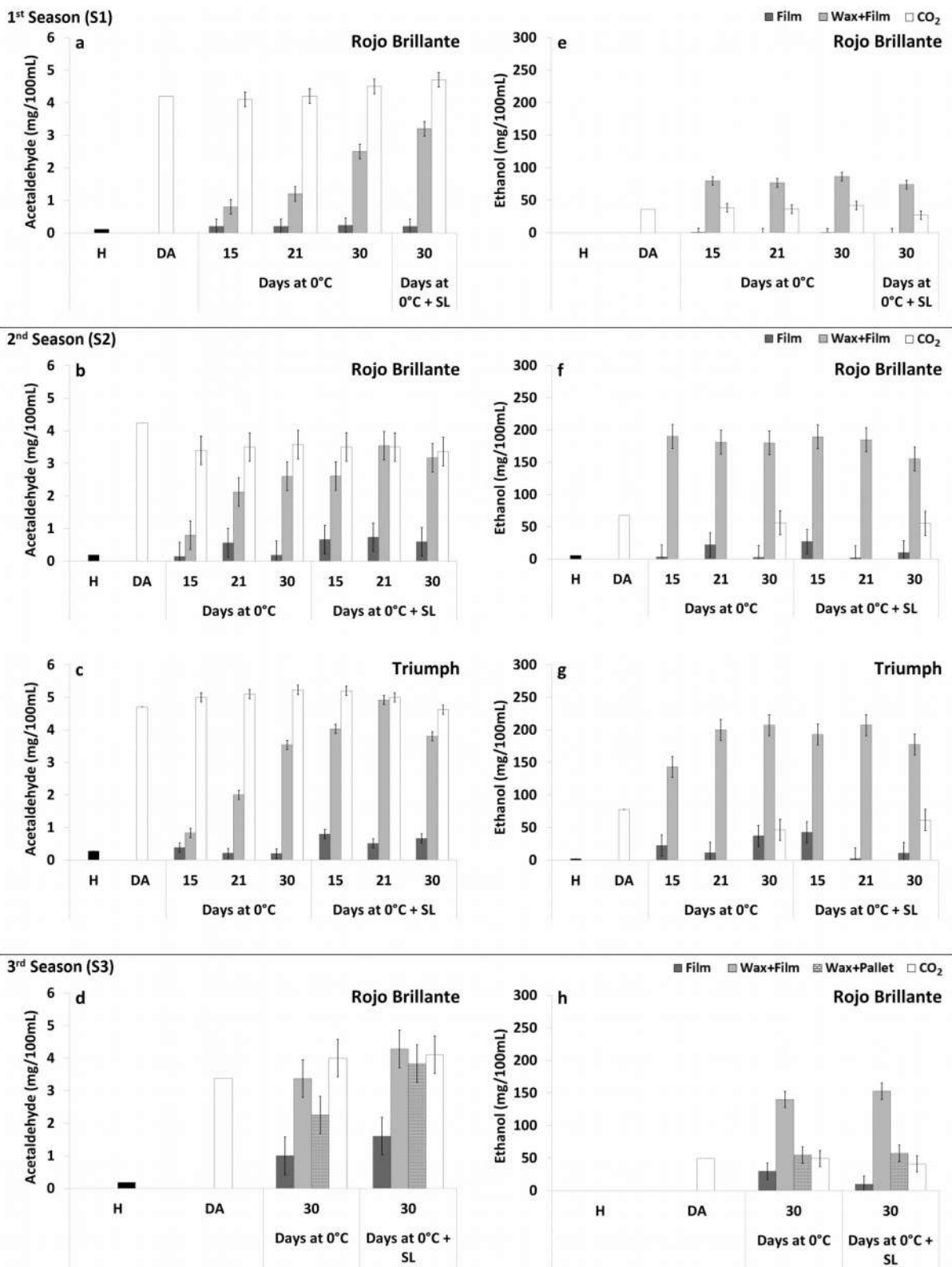


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529**Fig. 1** Soluble tannins content (a, b, c, d) and sensory astringency evaluation (e, f, g, h) of
 530persimmons ‘Rojo Brillante’ and ‘Triumph’ at harvest (H) during storage at 0°C and after the
 531subsequent 5-day shelf-life period at 20°C in season 1 (S1), season 2 (S2) and season 3 (S3).
 532Vertical bars represent LSD intervals ($p \leq 0.05$). Sensory astringency evaluation was from 1

533= no astringency to 5 = astringency; a value below 1.5 is considered commercially
534acceptable.

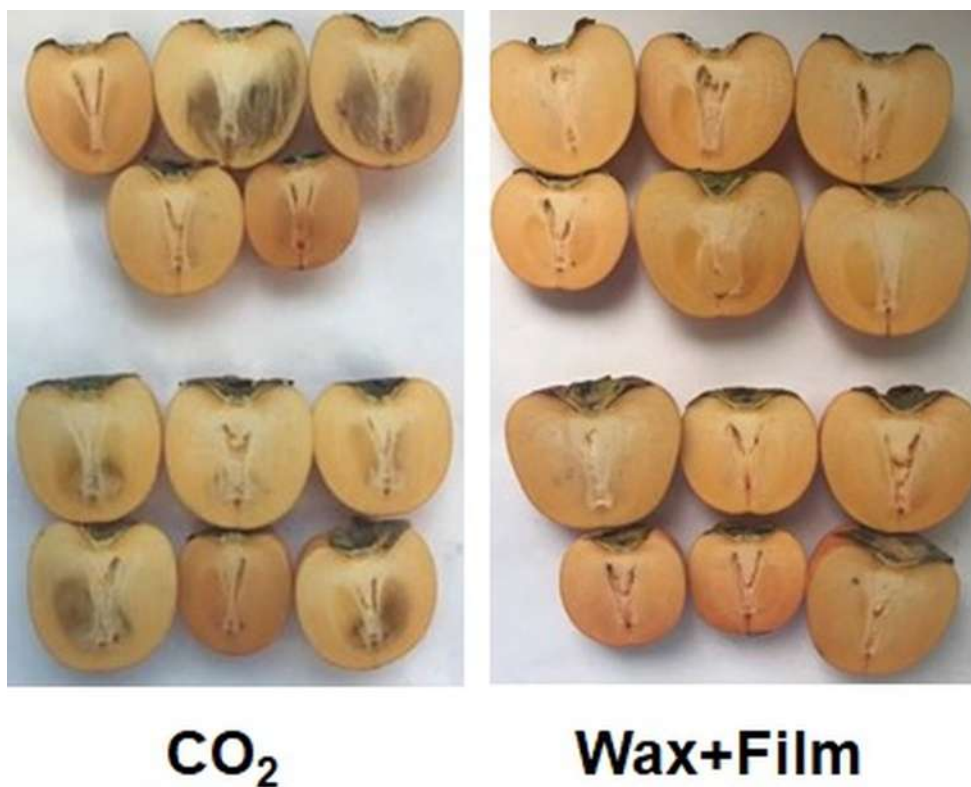
535Film: fruit packed in plastic film without wax. Wax+Film: fruit waxed and packed in plastic
536film. CO₂: fruit submitted to CO₂ treatment. Wax+Pallet: fruit waxed and boxes stacked on a
537wooden pallet which was strapped with macroperforated stretch film. DA: 1 day after CO₂
538treatment.



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540**Fig. 2** Acetaldehyde (a, b, c, d) and ethanol (e, f, g, h) (mg/100 mL) of persimmons ‘Rojo
 541Brillante’ and ‘Triumph’ at harvest (H), during storage at 0°C and after the subsequent 5-day
 542shelf-life period at 20°C in season 1 (S1), season 2 (S2) and season 3 (S3). Vertical bars
 543represent LSD intervals ($p \leq 0.05$).

544Film: fruit packed in plastic film without wax. Wax+Film: fruit waxed and packed in plastic
545film. CO₂: fruit submitted to CO₂ treatment. Wax+Pallet: fruit waxed and boxes stacked on a
546wooden pallet, which was strapped with macroperforated stretch film. DA: 1 day after CO₂
547treatment.



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550 **Fig. 3** Internal quality of the 'Rojo Brillante' fruit treated with either CO₂ or the new wax
551 after 30 days of cold storage, plus the 5-day shelf-life at 20 °C

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562**Table 1**

563Treatments and conditions tested for ‘Rojo Brillante’ and Triumph’ in the three season
564studied

Season	Treatments	Cultivar	Cold storage (CS) (0 °C)			Shelf-life (5d at 20 °C after CS)		
			15d	21d	30d	15d	21d	30d
S1	CO ₂ Wax+Film Film	Rojo Brillante	X	X	X	-	-	X
S2	CO ₂ Wax+Film Film	Rojo Brillante Triumph	X	X	X	X	X	X
S3	CO ₂ Wax+Film Film Wax+Pallet	Rojo Brillante	-	-	X	-	-	X

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577**Table 2**

578EtOH ($\mu\text{L/L}$) and CO_2 (%) of persimmons 'Rojo Brillante' and 'Triumph' during storage at 0
579°C in season 1 (S1), season 2 (S2) and season 3 (S3).

580*Different letters indicate statistical differences ($p \leq 0.05$).

581Film: fruit packed in plastic film without wax. Wax+Film: fruit waxed and packed in plastic

582film

583

		Days at 0 °C							
Season (Cultivar)	Treatment	1d		15d		21d		30d	
		EtOH	CO_2	EtOH	CO_2	EtOH	CO_2	EtOH	CO_2
S1	Film	-	-	0a*	0.2b	0a	0.6b	0	0a
(Rojo Brillante)	Wax+Film	-	-	0.02b	0.4b	0.02b	0.7b	0.02b	0.01a
S2	Film	0a	0.63b	0.01a	0.32a	0.01a	0.69b	0.01a	0.36a
(Rojo Brillante)	Wax+Film	0.67d	0.6b	0.08c	0.66b	0.04b	0.57b	0.02a	0.53b
S2	Film	0a	0.36a	0.01a	0.72b	0.01a	0.72b	0.01a	0.42a
(Triumph)	Wax+Film	0.47c	0.49a	0.04b	0.84b	0.04b	0.84b	0.02a	0.51a
S3	Film	-	-	-	-	-	-	0a	-
(Rojo Brillante)	Wax+Film	-	-	-	-	-	-	0.02a	-

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