

Document downloaded from:

[\[http://redivia.gva.es/handle/20.500.11939/7157\]](http://redivia.gva.es/handle/20.500.11939/7157)

This paper must be cited as:

[Palomo, J. L., Shima, M., Monterde, A., Navarro, I., Barbé, S., & Marco-Noales, E. First report of bacterial leaf blight caused by *Xanthomonas hortorum* pv. *carotae* on carrots in Spain. Plant disease. Published on-line, 23 february 2021.]

**ivia**  
Institut Valencià  
d'Investigacions Agràries

The final publication is available at

[\[https://doi.org/10.1094/PDIS-11-20-2493-PDN\]](https://doi.org/10.1094/PDIS-11-20-2493-PDN)

Copyright [APS]

## **First report of bacterial leaf blight caused by *Xanthomonas hortorum* pv. *carotae* on carrots in Spain**

J. L. Palomo,<sup>1</sup> M. Shima,<sup>1</sup> A. Monterde,<sup>2</sup> I. Navarro,<sup>2</sup> S. Barbé,<sup>2</sup> and E. Marco-Noales,<sup>2,†</sup>

<sup>1</sup> Centro Regional de Diagnóstico, Junta de Castilla y León, Salamanca, Spain

<sup>2</sup> Instituto Valenciano de Investigaciones Agrarias (IVIA), Valencia, Spain

† Corresponding author: [marco\\_est@gva.es](mailto:marco_est@gva.es)

In September 2019, symptoms resembling those of bacterial leaf blight were observed on carrot plants (*Daucus carota* L. subsp. *sativus* Hoffm.) cv. Romance cultivated in commercial plots in Chañe (Segovia), Spain. Symptoms were observed in two plots surveyed representing three hectares, with an incidence greater than 90%, and also in some plots in other nearby municipalities sown with the same batch of seeds. The lesions observed at the ends of the leaves were initially yellow that develop dark brown to black with chlorotic halos on leaflets that turned necrotic. Yellow, *Xanthomonas*-like colonies were isolated onto YPGA medium (Ridé 1969) from leaf lesions. Two bacterial isolates were selected and confirmed by real-time PCR using a specific primer set for *Xanthomonas hortorum* pv. *carotae* (Temple et al. 2013). All isolates were gram-negative, aerobic rods positive for catalase, able of hydrolyzing casein and aesculin and growing at 2% NaCl, while were negative for oxidase and urease tests. Sequences of 16S rRNA gene showed 100% similarity with *Xanthomonas campestris*, *X. arboricola*, *X. gardneri*, *X. cynarae* strains (GenBank accession numbers: MW077507.1 and MW077508.1 for the isolates CRD19-206.3 and CRD19-206.4, respectively). However, the resulting phylogeny of multilocus sequence analysis (MLSA) of a concatenation of the housekeeping genes *atpD*, *dnaK*, and *efp* (Bui Thi Ngoc et al. 2010), by using neighbour-joining trees generated with 500 bootstrap replicates, grouped the two isolates with the *X. hortorum* pv. *carotae* M081 strain (Kimbrel et al. 2011) (GenBank accession numbers: MW161270 and MW161271 for *atpD* for the two isolates, respectively; MW161268 and MW161269 for *dnaK*; MW161272 and MW161273 for *efp*). A pairwise identity analysis revealed a 100% identity between all three isolates. Pathogenicity of the isolates was tested by spray inoculation (Christianson et al. 2015) with a bacterial suspension ( $10^8$  CFU/ml) prepared in sterile distilled water at 3 to 4 true-leaf stage (six plants per isolate). Sterile distilled water

was used as negative control. The inoculated plants were incubated in a growth chamber (25°C and 95% relative humidity [RH]) for 72 h, and then transferred to a greenhouse at 24 to 28°C and 65% RH. Characteristic leaf blight symptoms developed on inoculated carrot plants, while no symptoms were observed on the negative control plants 20 days after inoculation. The bacterium was re-isolated from symptomatic tissue and the identity confirmed through PCR analysis. Based on PCR, morphological and phenotypic tests, sequence analysis, and pathogenicity assays, the isolates were identified as *X. hortorum* pv. *carotae*. To our knowledge, this is the first report of bacterial leaf blight of carrot caused by *X. hortorum* pv. *carotae* in Spain, and the first molecular and pathological characterization. It is important to early detect this pathogen and take suitable measures to prevent its spread, since it could cause yield losses for a locally important crop such as carrot.

#### References:

- Bui Thi Ngoc, L., et al. 2010. Int. J. Syst. Evol. Microbiol. 60:515. doi: 10.1099/ijms.0.009514-0.
- Christianson, C. E., et al. 2015. HortScience 50:341. <https://doi.org/10.21273/HORTSCI.50.3.341>.
- Kimbrel J. A., et al. 2011. Mol. Plant. Pathol. 12:580. doi: 10.1111/j.1364-3703.2010.00694.x.
- Ridé, M. 1969. In: Bourgin CVM editor. Les Bactérioses et les Viroses des Arbres Fruitières. Ponsot, Paris, France.
- Temple, T. N., et al. 2013. Plant Dis. 97:1585. <https://doi.org/10.1094/PDIS-03-13-0262-RE>.



Fig. 1S. Symptoms of bacterial leaf blight observed on carrot plants in commercial plots in Chañe (Segovia, Spain). (A) Leaves presented dark brown to black coloration with chlorotic halos on leaflets that turned necrotic; (B) detail of the lesions.

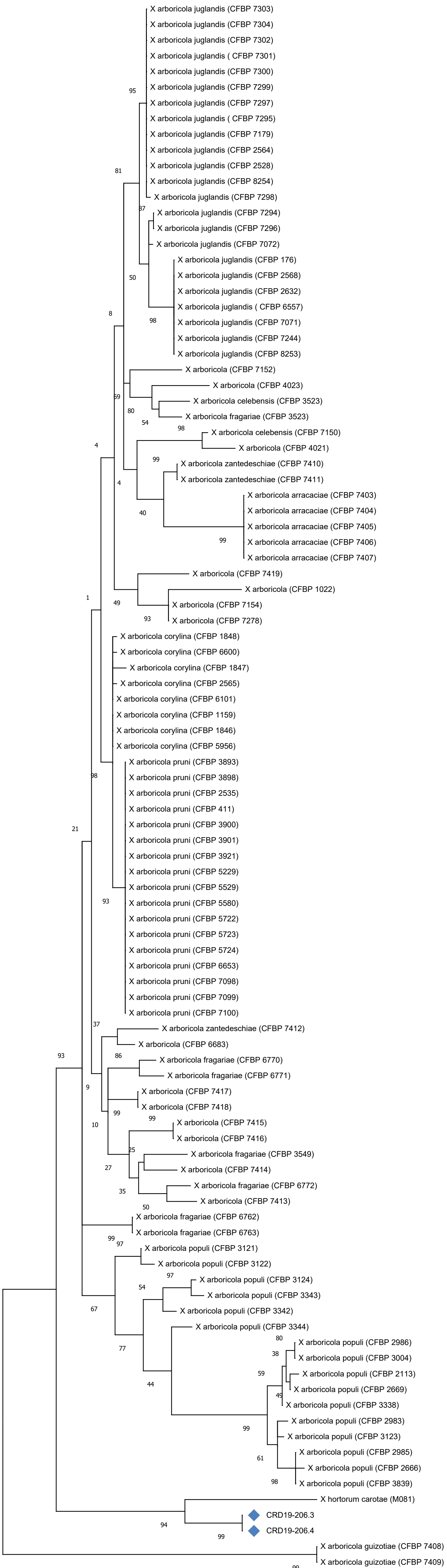


Fig. 2S. Complete neighbor joining tree of concatenated, nucleotide sequences for partial *atpD*, *dnaK*, and *efp* genes from 100 *Xanthomonas* strains (Kumar et al. 2018). Numbers indicate bootstrap support ( $r = 500$ ). Evolutionary analyses were conducted in MEGA X (Young et al 2018). The scale bars indicate the number of amino acid substitutions per site.

References:

- Kumar, S., et al. 2018. Mol. Biol Evol. 35:1547. doi: 10.1093/molbev/msy096.
- Young, J. M., et al. 2018. Syst. Appl. Microbiol. 31:366. doi: 10.1016/j.syapm.2008.06.004.