The role of horizontal gene transfer in *Staphylococcus aureus* host adaptation

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***S**taphylococcus aureus* **is an important human pathogen that also causes economically important infections of livestock. In a recent paper, we employed a population genomic approach to investigate the molecular basis of ruminant host adaptation by *S. aureus*. The data suggest that the common pathogenic clone associated with small ruminants originated in humans but has since adapted to its adopted host through a combination of allelic diversification, gene loss and acquisition of mobile genetic elements. In particular, a new subfamily of staphylococcal pathogenicity islands (SaPI) was identified encoding a novel von Willebrand factor-binding protein (vWBP) with ruminant-specific coagulase activity. The wide distribution of vWBP-encoding SaPIs among ruminant strains implies an important role in host-adaptation. In the current article we summarize the findings of the paper and comment on the implications of the study for our understanding of the molecular basis of bacterial host adaptation.**

Staphylococcus aureus is a notorious human pathogen that can also cause important diseases of animals including mastitis in ruminants,[1] dermatitis in rabbits[2] and skeletal infections in poultry.[3] Strains of *S. aureus* with combinations of phenotypes which were unique to different host species were first identified in the 1930s, leading to the description of host-specific ecological variants (ecovars).[4-6] Subsequently, numerous population genetic studies have demonstrated that the majority of *S. aureus* animal infections are caused by pathogenic clones which are not commonly found in association with humans, implying that they are host-specialized and largely host-restricted.[7-10] While the molecular basis of the adaptation of *S. aureus* to animal hosts has not been well examined, recent studies have identified genetic determinants that correlate with infection of a particular host.[11-13] For example, Lowder et al. provided the first clear evidence of a human-to-poultry host jump by a bacterial pathogen, which led to the emergence of an avian-adapted pandemic clone.[11,14] The origin of the clone was traced to a subtype of the common human clonal complex CC5 about 40 years ago but it has since undergone inter-continental dissemination due to the globalized nature of the poultry industry.[11] Of note, a large-scale acquisition of mobile genetic elements (MGE) from other *S. aureus* strains resident on birds was discovered, suggesting a key role for horizontal gene acquisition in avian host adaptation.[11] Furthermore, analysis of the genome sequence of a bovine mastitis strain RF122 of the ST151 clone of revealed novel MGE present in other bovine strains but absent in human isolates.[13]

Recently, we employed a combination of population genetics, comparative genomics and functional analysis to investigate the evolutionary origin of the major clonal complex affecting small ruminants (CC133).[14] The data suggest that the CC133 clone originated as a result of a host jump from humans to ruminants followed by genetic adaptation through

a combination of allelic diversification, genome decay and acquisition of mobile elements containing virulence genes with enhanced activity in ruminants.[14] In comparison to *S. aureus* strains of human origin, allelic variation was identified in genes involved in toxin production, metabolism, adherence, replication and repair, and regulatory pathways, including many which were under diversifying selective pressure.[14] Gene decay is a common feature of bacteria undergoing niche adaptation, and this has previously been observed among *S. aureus* clones adapted to cows and poultry.[12,13] Similarly, ED133 contained pseudogenes resulting in loss of function of putative virulence factors such as toxins and lipoproteins.[14] Of note, several pseudogenes were shared with the bovine clone ST151 but contained unique underlying mutations suggestive of a strong selective pressure for loss of function in ruminants.[12-14] The most striking feature of the genome of ED133 was a unique complement of MGE that had not been previously identified among strains from humans or other animals. In total, two new members of the *S. aureus* pathogenicity island (SaPI) family and three novel prophages were identified.[14] Of these, two of the prophages and both SaPIs were widely distributed among other isolates of the CC133 lineage but not among other ruminant, avian or human lineages examined, consistent with a function which is specific for the small ruminant niche occupied by CC133 strains.

The capacity of ruminant strains of *S. aureus* to coagulate bovine or ovine plasma has been employed as a component of the traditional biotyping scheme used to differentiate *S. aureus* host-specific ecovars.[6] However, the molecular basis for this phenotype was previously unknown. A novel SaPI (SaPIov2), widespread among CC133 strains encoded a unique variant of the core genome-encoded von-willebrand factor binding protein (vWBp[Sov2]).[14] Similar to the chromosomally encoded vWBp, vWBp[Sov2] contained a predicted coagulase domain which has previously been implicated in *S. aureus* pathogenesis.[15] We speculated that SaPIov2 may confer the capacity to coagulate ruminant plasma and constructed isogenic CC133 ruminant strains, differing only in the presence

of SaPIov2. In each case, the presence of SaPIov2 conferred the ability to coagulate ruminant plasma, revealing the basis for a defining phenotype of the *S. aureus* biotyping scheme.[14] Independently, Viana et al. identified 3 additional SaPIs encoding variant vWBPs among strains of bovine and equine origin, and the authors demonstrated that the plasma coagulation phenotype was due to the activity of the encoded vWBP.[16] It appears that diversification of the N-terminal coagulase domain of vWBp has resulted in the capacity to activate equine or ruminant prothrombin leading to the conversion of soluble fibrinogen to insoluble fibrin.[16] Of note, an equine strain of the same clonal complex as ovine strain ED133 (CC133) contained a SaPI encoding a vWBP with the capacity to coagulate equine plasma, consistent with a central role for this family of SaPIs in the host-tropism of *S. aureus*. Recently, Cheng et al. demonstrated the importance of the archetypal *S. aureus* coagulase and the chromosomal copy of vWBP in the formation of abscesses in a mouse model.[15] The role of plasma coagulase activity in ruminant mastitis is unclear but we speculate that it may contribute to the formation of micro-abscesses during intramammary infection, which may facilitate persistence during chronic infection.

An understanding of the timescale for the evolution of livestock-associated *S. aureus* is important to appreciate the ecological context of their emergence and the capacity for new pathogenic clones affecting animals to evolve. To predict the date of the putative human-to-ruminant host jump that led to the CC133 clonal complex, a relaxed molecular clock based method was employed with the program BEAST.[14,17] The estimated host switch was calculated to have occurred about 115 to 1204 years ago, based on a mutation rate of 3.3×10^{-6} per site per year calculated for contemporary isolates of the important human clone ST239.[18] However, the relevance of this mutation rate for predicting long term evolutionary events is unclear as the effects of purifying selection may limit the applicability of mutation rates calculated over shorter timescales. Furthermore, our current appreciation of the variation in mutation rates for different *S. aureus* clones occupying diverse niches is very

limited. It is reasonable to assume that our prediction is the minimum date a host switch could have happened but it may have been much earlier. We predict that it is likely to have happened since the domestication of small ruminants (approximately ~11,000 years ago[19]), which would have resulted in ample opportunities for the transfer of bacteria between human and livestock. However, these important questions remain to be answered.

Overall, this population genomics study and other related works have improved our understanding of the evolutionary origin of livestock-associated *S. aureus* and identified some of the genetic determinants that differentiate animal strains from human pathogenic clones.[11,13,14,16] In particular, although possibly not surprising, MGE appear to play a fundamental role in facilitating the adaptation of *S. aureus* to different host species. The origin of the MGE is unclear but it is likely that other staphylococcal species that are normally resident on animal hosts may represent the source. It is feasible that the functional characterization of the role of the identified MGE in bacterial adaptation to animal hosts may lead to the design of novel approaches for controlling livestock infections.

References

1. Bergonier D, de Cremoux R, Rupp R, Lagriffoul G, Berthelot X. Mastitis of dairy small ruminants. Vet Res 2003; 34:689-716.
2. Hermans K, Devriese LA, Haesebrouck F. Rabbit staphylococcosis: difficult solutions for serious problems. Vet Microbiol 2003; 91:57-64.
3. McNamee PT, McCullagh JJ, Thorp BH, Ball HJ, Graham D, McCullough SJ, et al. Study of leg weakness in two commercial broiler flocks. Vet Rec 1998; 143:131-5.
4. Madison RR. Fibrinolytic staphylococci. Proc Sac Exp Biol 1935; 33:309.
5. Minnett FC. Staphylococci from animals with particular reference to toxin production. J Pathol Bacteriol 1936; 42:247-59.
6. Devriese LA. A simplified system for biotyping *Staphylococcus aureus* strains isolated from animal species. J Appl Bacteriol 1984; 56:215-20.
7. Kapur V, Sischo WM, Greer RS, Whittam TS, Musser JM. Molecular population genetic analysis of *Staphylococcus aureus* recovered from cows. J Clin Microbiol 1995; 33:376-80.
8. Fitzgerald JR, Meaney WJ, Hartigan PJ, Smyth CJ, Kapur V. Fine-structure molecular epidemiological analysis of *Staphylococcus aureus* recovered from cows. Epidemiol Infect 1997; 119:261-9.

9. Smith EM, Green LE, Medley GF, Bird HE, Fox LK, Schukken YH, et al. Multilocus sequence typing of intercontinental bovine *Staphylococcus aureus* isolates. J Clin Microbiol 2005; 43:4737-43.

10. Jorgensen HJ, Mork T, Hogasen HR, Rorvik LM. Enterotoxigenic *Staphylococcus aureus* in bulk milk in Norway. J Appl Microbiol 2005; 99:158-66.

11. Lowder BV, Guinane CM, Ben Zakour NL, Weinert LA, Conway-Morris A, Cartwright RA, et al. Recent human-to-poultry host jump, adaptation and pandemic spread of *Staphylococcus aureus*. Proc Natl Acad Sci USA 2009; 106:19545-50

12. Herron LL, Chakravarty R, Dwan C, Fitzgerald JR, Musser JM, Retzel E, et al. Genome sequence survey identifies unique sequences and key virulence genes with unusual rates of amino Acid substitution in bovine *Staphylococcus aureus*. Infect Immun 2002; 70:3978-81.

13. Herron-Olson L, Fitzgerald JR, Musser JM, Kapur V. Molecular correlates of host specialization in *Staphylococcus aureus*. PLoS One 2007; 2:1120.

14. Guinane CM, Ben Zakour NL, Tormo-Mas MA, Weinert LA, Lowder BV, Cartwright RA, et al. Evolutionary genomics of *Staphylococcus aureus* reveals insights into the origin and molecular basis of ruminant host adaptation. Genome Biol Evol 2010; 2:454-66.

15. Cheng AG, McAdow M, Kim HK, Bae T, Missiakas DM, Schneewind O. Contribution of coagulases towards *Staphylococcus aureus* disease and protective immunity. PLoS Pathog 2010; 6:1001036.

16. Viana D, Blanco J, Tormo-Mas MA, Selva L, Guinane CM, Baselga R, et al. Adaptation of *Staphylococcus aureus* to ruminant and equine hosts involves SaPI-carried variants of von Willebrand factor-binding protein. Mol Microbiol 2010; 77:1583-94.

17. Drummond AJ, Rambaut A. BEAST: Bayesian evolutionary analysis by sampling trees. BMC Evol Biol 2007; 7:214.

18. Harris SR, Feil EJ, Holden MT, Quail MA, Nickerson EK, Chantratita N, et al. Evolution of MRSA during hospital transmission and intercontinental spread. Science 2010; 327:469-74.

19. Zeder MA. Domestication and early agriculture in the Mediterranean Basin: Origins, diffusion and impact. Proc Natl Acad Sci USA 2008; 105:11597-604.