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**1Effect of ethylene degreening on the development of postharvest penicillium molds
2and fruit quality of early season citrus fruits**

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24Abstract

25The effect of commercial degreening with ethylene gas on fruit susceptibility and quality
26and development of postharvest green (GM) and blue (BM) molds on early season citrus
27fruits was investigated. Each cultivar was harvested with different peel color indexes (CI).
28Fruit were exposed for 3 d to 2 $\mu\text{L L}^{-1}$ ethylene at 21°C and 95-100% RH before or after
29artificial inoculation with *Penicillium digitatum* or *P. italicum*. Control fruit were kept at
30the same environmental conditions without ethylene. Fruit were stored at either 20 °C for 7
31d or 5 °C for 14 d and disease incidence (%) and severity (lesion diameter) were assessed.
32No significant effect of commercial degreening was observed on fruit susceptibility to both
33GM and BM on citrus cultivars inoculated after degreening. Likewise, no significant effect
34was observed on disease incidence on citrus cultivars inoculated before degreening and
35stored at either 20 °C for 7 d or 5 °C for 14 d. In contrast, in cultivars like ‘Clemenules’
36mandarins and ‘Navelina’ oranges, degreening significantly increased the severity on fruit
37with higher initial CI (-3.6 and 1.7, respectively). GM and BM severity on degreened and
38control ‘Clemenules’ mandarins incubated at 20 °C for 7 d was 146 and 118 mm and 56
39and 46 mm, respectively. In general, commercial degreening did not significantly affect
40external and internal quality attributes of citrus cultivars. Commercial degreening after
41inoculation of less green (more mature) fruit showed a trend to increase mold severity,
42presumably through an aging effect (acceleration of peel senescence).

43**Keywords:** Orange, mandarin, postharvest disease, *Penicillium digitatum*, *P. italicum*, peel
44 color index, fruit quality

45

461. Introduction

47 Postharvest green mold (GM), caused by *Penicillium digitatum* (Pers.:Fr.) Sacc., and
48 postharvest blue mold (BM), caused by *Penicillium italicum* Wehmer, are the most
49 economically important postharvest diseases of citrus in Spain and all production areas
50 characterized by low summer rainfall (Eckert and Eaks, 1989). Blue mold is especially
51 important on citrus fruits kept under cold storage (Palou et al., 2001).

52 External color of citrus fruits is more related to climatic conditions than to internal
53 maturity. Degreening with ethylene (C₂H₄) is a common commercial practice employed in
54 many parts of the world to accelerate artificially the natural color break in which fruit turns
55 from green to orange/yellow (Porat, 2008; Peng et al., 2013). Particularly, it is extensively
56 implemented in Spain from September to December to market early season mandarins and
57 oranges. Since 1970's decade to date, several research works have been conducted to study
58 the relationship between ethylene applications and postharvest decay by phytopathogenic
59 fungi on citrus fruit, finding a non-well-determined trend. Discrepancies observed in
60 research results seem to be related to different environmental conditions, the amount of
61 ethylene applied and/or different host-pathogen interactions. On the one hand, fruit
62 degreening has been reported to induce resistance to penicillium molds or anthracnose
63 caused by *Colletotrichum gloeosporioides*, especially when applied at 30 °C and relative
64 humidity (RH) higher than 90% (Brown, 1973, 1975, El-Kazzaz et al., 1983a; Porat et al.,
65 1999). On the other hand, other research results have suggested an increase in the incidence
66 of citrus postharvest diseases such as stem-end rots or penicillium molds following ethylene
67 exposure (McCornack, 1971; Barmore and Brown, 1985; Brown, 1986; Zhang, 2004). It
68 was reported in another work that degreening in standard conditions at 20 °C after fungal

69 inoculation had no effect on the incidence of both GM and BM on ‘Clemenules’ mandarin
70 fruit (Plaza et al., 2004). Given these different degreening effects depending on specific
71 handling and environmental conditions, it is important to establish them for each particular
72 situation. Standard commercial degreening practices for early season mandarins and
73 oranges in Spain, California and other Mediterranean-climate areas consist of fruit exposure
74 for 2-4 d to 2-5 $\mu\text{L L}^{-1}$ ethylene at 20-22 °C and RH > 90% (Sdiri et al., 2012a), while in
75 other citrus producing areas like Florida or Brazil, commercial degreening is typically
76 performed at temperatures surrounding 30 °C (Brown, 1973, 1975).

77 Regarding the effects of ethylene exposure on the quality of harvested horticultural
78 produce, the responses are numerous and varied, and they can be beneficial or detrimental
79 depending on each case (Saltveit, 1999; Palou et al., 2003). In the case of citrus fruit, it is
80 important to examine the effects of ethylene degreening on fruit quality attributes other
81 than peel color, especially on fruit destined to prolonged storage or long-distance markets.
82 Thus, the effects of ethylene or 1-methylcyclopropene (1-MCP), an ethylene action
83 inhibitor, on fruit weight loss, firmness, total soluble solids content (SSC), titratable acidity
84 (TA), or consumer acceptance for different citrus cultivars have been assessed with
85 different results (Porat et al., 1999; Plaza et al., 2004; Tietel et al., 2010; Mayuoni et al.,
86 2011a). Furthermore, the knowledge of the effects on nutritional quality and content of
87 bioactive compounds is increasingly gaining importance (Mayuoni et al., 2011b; Sdiri et
88 al., 2012b).

89 Before the interest of the Spanish citrus industry to optimize degreening treatments and
90 overall fruit handling in the packinghouses for the most representative commercial
91 cultivars, it is important to determine the influence of ethylene degreening in our particular

92 conditions on the development of GM and BM, the most economically important cause of
93 postharvest decay, and also on fruit susceptibility to these diseases. Therefore, the aims of
94 this research were to: (i) determine the effect of commercial degreening with ethylene gas
95 on the susceptibility to GM and BM of intact early season mandarins and oranges (ii) assess
96 the effect of degreening on the incidence and development of GM and BM on fruit
97 previously inoculated with *P. digitatum* or *P. italicum* and incubated at 20 °C or stored at 5
98 °C, and (iii) study the effect of degreening at standard commercial conditions on internal
99 and external fruit quality attributes. Preliminary results from this research have been
100 recently published (Moscoso-Ramírez and Palou, 2013).

101

1022. Materials and methods

1032.1. Fruit

104 The trials were conducted from 2008 to 2011 with ‘Clemenpons’ and ‘Clemenules’
105 clementine mandarins (*Citrus reticulata* Blanco), ‘Navelina’ oranges (*Citrus sinensis* (L.)
106 Osbeck), and ‘Nova’ [*C. reticulata* x (*Citrus reticulata* x (*Citrus reticulata* x *Citrus*
107 *paradisi*)), synonym: ‘Clemenvilla’] hybrid mandarins. Fruit were collected from
108 commercial orchards in the Valencia area (Spain) and used the same day or stored up to 1
109 week at 5 °C and 90% RH before use. Before each experiment, fruit were selected,
110 randomized, washed, disinfected superficially by immersion for 2 min in a 0.5% sodium
111 hypochlorite solution, rinsed with tap water to eliminate residual chlorine, and allowed to air
112 dry at room temperature. Depending on the experiment, fruit of each cultivar were
113 harvested at different rind color indexes (CI = -0.07, 0.9 for ‘Clemenpons’ mandarins; CI =
114 -6.5, -3.6, 2.2 for ‘Clemenules’ mandarins; CI = -5.3, 1.1, 1.7 for ‘Navelina’ oranges; CI =

11512.3 for ‘Nova’ hybrid mandarins), in some cases during the same season and in other cases
116in different seasons.

117

1182.2. Commercial degreening procedure

119 Fruit were transported to a 1000 t commercial degreening room in a local citrus
120packinghouse (Fontestad S.A., Montcada, Valencia). Fruit were exposed to 2 $\mu\text{L L}^{-1}$
121ethylene ($\pm 0.5 \mu\text{L L}^{-1}$) at a constant temperature of 21 °C ($\pm 0.5 \text{ }^\circ\text{C}$) and RH > 95% for 72
122h, with the exception of fruit from one of the experiments with ‘Clemenules’ mandarins
123that needed 24 h of additional exposure to develop acceptable commercial color. At the
124same time, non-degreened (control) fruit were exposed to the same environmental
125conditions but without exposure to exogenous ethylene in a storage room located in the
126IVIA CTP pilot plant.

127

1282.3. Fungal inoculation

129 *Penicillium digitatum* and *P. italicum*, isolates NAV-7 and MAV-1, respectively,
130from the fungal culture collection of the IVIA CTP, were cultured on potato dextrose agar
131(PDA, Sigma-Aldrich Chemical Co., St. Louis, MA, USA) plates at 25 °C. Conidia of each
132fungus from 7-to 14-day-old were taken from the plate surface with a sterile glass rod and
133transferred to a sterile aqueous solution of 0.05% Tween 80® (Panreac, S.A.U., Barcelona,
134Spain). Conidial suspensions were filtered through two layers of cheesecloth to separate
135hyphal fragments and adjusted to a concentration of 1×10^5 spores mL^{-1} using a
136haemocytometer. Fruit were wounded and inoculated with the pathogens at the same time
137by immersing the tip of a stainless steel rod (2 mm length and 1 mm diameter) into the

138 conidial suspension and making a puncture on the peel in the equatorial region of the fruit.

139 Different lots of fruit were inoculated with each fungus.

140

141 2.4. Effect on fruit susceptibility to disease

142 To evaluate the effect of degreening on the fruit susceptibility to GM and BM, fruit were
143 artificially inoculated with *P. digitatum* or *P. italicum* about 2 h after ethylene degreening.

144 This set of experiments was performed with fruit of each cultivar harvested at the lowest CI
145 (CI of -0.07, -6.5, -5.3 and 12.3 for ‘Clemenpons’, ‘Clemenules’, ‘Navelina’ and ‘Nova’
146 cultivars). Each treatment consisted of 4 replications with 10 fruit each. Degreened and
147 non-degreened inoculated fruit were incubated at 20°C and 90% RH for 7 d. Fruit were
148 examined after 3 and 7 d to determine the incidence (% of infected wounds) and severity
149 (diameter of lesion in mm measured only in infected fruit) of the molds.

150

151 2.5. Effect on disease development

152 To evaluate the effect of degreening on fungal development, fruit were inoculated with
153 the pathogens as previously described 2 h before ethylene degreening. For each cultivar,
154 these experiments were performed with fruit harvested at all different CI previously
155 mentioned. After commercial degreening, degreened and control fruit were stored under
156 two different conditions: (i) incubation at 20 °C and 90% RH for 7 d, and (ii) cold storage
157 at 5 °C and 90% RH for 2 weeks. Stored fruit were periodically examined to determine
158 disease incidence and severity. For each pathogen, treatment and storage condition, 4
159 replications of 10 fruit each were used.

160

1612.6. Effect on citrus fruit quality

162 Non-inoculated oranges and mandarins were used for quality assessment. Fruit external and
163 internal quality were determined just after harvest (initial quality) and 2-3 d after
164 degreening on degreened and non-degreened (control) fruit (final quality).

1652.6.1. External quality

166 Peel color was measured using Hunter parameters (L, a, b) with a colorimeter (Model
167 Minolta CR-300, Konica Minolta Business Technologies, Inc., Tokio, Japan). A color
168 index (CI) was calculated: $CI = 1000.a/L.b$ (Jiménez-Cuesta et al., 1981). For each
169 treatment, three measurements on the equatorial area of 25 fruit were performed.

170 Firmness of 20 fruit per treatment was determined using an Instron Universal Testing
171 Machine (Model 4301, Instron Corp., Norwood, MA, USA). Each fruit was compressed
172 between two flat surfaces closing together at the rate of 5 mm min^{-1} . The machine gave the
173 deformation (mm) after application of a load of 1 kg to the equatorial region of the fruit.
174 Results were expressed as percentage of deformation related to initial diameter. Peel break
175 resistance was measured on 30 fruit per treatment using the same machine. Each fruit was
176 compressed with a 5 mm diameter steel rod until the fruit rind was broken, and the
177 necessary pressure (kg) was measured.

178 Peel oil release pressure (kg) of 20 fruit per treatment was determined using a fruit
179 pressure tester with a 8 mm diameter tip (Model FT327, Facchini, Alfonsine, Italy). Each
180 fruit was wrapped with filter paper and then compressed with the tester until essential oil
181 stains appeared. The necessary pressure (kg) was annotated.

1822.6.2. Internal quality

183 The fruit internal quality was only determined in clementine mandarins (cvs.
184 'Clemenpons' and 'Clemenules'). The juice from 3 previously weighed samples of 8 fruit

185each was extracted with a rotatory citrus squeezer and filtered through a 0.8 mm diameter
186sieve. The following fruit internal quality parameters were determined: SSC was measured
187with a digital refractometer (Model DR-101, Optic Ivymen System, Barcelona, Spain) and
188expressed as percentage. TA was determined from a 5 mL aliquot by titration with 0.1 N
189NaOH with phenolphthalein indicator and results were given as g of citric acid per 100 mL
190(%). MI was calculated as the SSC/TA ratio. In all cases, two replicated measures were
191performed with each juice sample. Juice yield was expressed as percentage of juice (mL)
192per fruit weight (g).

193

1942.7. Statistical analysis

195 Data from disease assessment and fruit quality parameters were analyzed by analysis of
196variance (ANOVA) with Statgraphics software (Statgraphics Plus, version 5.1). Data on
197disease incidence were transformed to the arcsine of the square root of the proportion of
198infected fruit to assure the homogeneity of variances. Statistical significance was judged at
199the level $P \leq 0.05$. When appropriated, the Fisher's Protected Least Significant Difference
200(LSD) test was applied to separate means. Shown values are non-transformed data.

201

2023. Results

2033.1. Effect of ethylene degreening on fruit susceptibility to disease

204 No significant effect of commercial degreening was observed on the incidence of
205both GM and BM on 'Clemenpons', 'Clemenules', 'Navelina' and 'Nova' cultivars
206harvested with initial peel CI of -0.07, -6.5, -5.3 and 12.3, respectively, artificially
207inoculated 2 h after degreening with *P. digitatum* or *P. italicum* and incubated at 20 °C and

20895-100% RH for 7 d. Similarly, the severity of the molds was not affected by commercial
209degreening (data not shown).

210

211**3.2.** Effect of ethylene degreening on disease development in fruit previously inoculated
212 with *P. digitatum* or *P. italicum*

213No significant effect of commercial degreening was observed on the incidence of both GM
214and BM on ‘Clemenpons’ (initial CI of -0.07 and 0.9; Fig. 1), ‘Clemenules’ (initial CI of -
2156.5, -3.6, and 2.2; Fig. 3) and ‘Nova’ (initial CI of 12.3; data not shown) mandarins, and
216‘Navelina’ oranges (initial CI of -5.3, 1.1, and 1.7; Fig. 5) inoculated with *P. digitatum* or
217*P. italicum* about 2 h before degreening and incubated at 20 °C and 90% RH for 7 d.
218Likewise, no effect was found on the incidence of both molds on ‘Clemenpons’,
219‘Clemenules’ and ‘Nova’ mandarins, and ‘Navelina’ oranges stored at 5 °C for 14 d, with
220the exception of ‘Clemenules’ mandarins with initial CI of -3.6, in which GM incidence on
221degreened and non-degreened fruit was 100 and 85%, respectively (Fig. 3); and ‘Navelina’
222oranges with initial CI of -5.3 and 1.7, in which GM incidence on degreened and non-
223degreened fruit was 90 and 63% and 98 and 70%, respectively (Fig. 5).

224 On ‘Clemenpons’ mandarins with the initial CI of -0.07 and 0.9, commercial
225degreening 2 h after fungal inoculation had no significant effect on the severity of the
226molds on fruit incubated at 20 °C for 7 d (Fig. 2). On ‘Clemenpons’ mandarins cold-stored
227at 5 °C for 14 d, commercial degreening did not significantly affect the severity of the
228molds with the exception of BM on mandarins with an initial CI of 0.9, in which lesion
229diameters were 33 and 27 mm on degreened and control fruit, respectively (Fig. 2).

230 On ‘Clemenules’ mandarins, the effect of commercial ethylene degreening on
231disease severity was dependent on the initial peel CI. Degreening treatment significantly

232increased the severity of the molds on fruit incubated at 20 °C for 7 d, with the exception of
233mandarins with an initial CI of -6.5 (Fig. 4). On mandarins cold-stored at 5 °C for 14 d,
234commercial degreening significantly increased the severity of the molds on mandarins with
235an initial CI of -3.6, with GM and BM severity values of 49 and 34 mm and 28 and 19 mm
236on degreened and non-degreened fruit, respectively. No significant effect was observed on
237fruit with an initial CI of -6.5 (Fig. 4).

238 On ‘Navelina’ oranges, commercial degreening had a significant effect on the
239severity of the molds, and it was dependent on the initial peel CI. Degreening treatment
240significantly increased the severity of the molds on fruit incubated at 20 °C for 7 d, with the
241exception of mandarins with an initial CI of -5.3 for both molds and of 1.1 for BM (Fig. 6).
242On mandarins cold-stored at 5 °C for 14 d, commercial degreening significantly increased
243the severity of the molds on mandarins with all three initial peel CI (-5.3, 1.1, 1.7), with the
244exception of mandarins with an initial peel CI of -5.3 for BM (Fig. 6).

245 On ‘Nova’ hybrid mandarins degreened with an initial peel CI of 12.3, ethylene
246degreening did not significantly affect the severity of the molds stored neither at 20 °C for 7
247d nor at 5 °C for 14 d, with the exception of GM severity, in which, lesion diameters were
24852 and 37 mm on degreened and control fruit, respectively (data not shown).

249

2503.3. Effect of ethylene degreening on citrus fruit quality

251In general, the external quality attributes of citrus cultivars with different initial peel CI
252were not influenced by ethylene degreening. Although significant differences between
253degreened and control fruit were found in few cases, the practical impact of such
254differences was minimal (Table 1). For instance, ‘Clemenules’ mandarin fruit degreened
255with an initial peel CI of -6.5 were significantly more firm (lower deformation) than non-

256degreened fruit, but the deformation values were 5.48 and 6.02%, respectively. Peel break
257resistance was significantly higher on degreened ‘Clemenules’ mandarins with an initial
258peel color index of -6.5 (1.7 kg) than on control fruit (1.3 kg), but the force difference was
259only of 0.4 kg (Table 1). Peel oil release pressure was significantly lower on degreened
260‘Clemenpons’ mandarins and ‘Navelina’ oranges, with the initial peel CI of 0.9 (3.99 kg)
261and -5.3 (5.40 kg), respectively, than on control fruit (4.93 and 5.40 kg, respectively).
262No significant effect of commercial degreening was observed on internal quality attributes
263(TA, SSC, MI, and juice yield) of ‘Clemenpons’ and ‘Clemenules’ clementine mandarins
264(Table 2).

265

2664. Discussion

267This research work has determined the effect of commercial degreening with ethylene gas
268at the standard Spanish conditions on fruit susceptibility and development of GM and BM
269in early season commercially valuable mandarins and oranges, as well as on the most
270important citrus fruit quality attributes. In general, commercial degreening before fungal
271inoculation with *P. digitatum* or *P. italicum* had no significant effect on the susceptibility to
272postharvest GM and BM on mandarins and oranges harvested with low CI and incubated at
27320 °C for 7 d. The implications of these results for the Spanish citrus industry are positive
274because, not being more susceptible, degreened fruit will not require special or additional
275preventive antifungal treatments to protect the fruit from infections by *Penicillium* spp. that
276may take place after degreening in the packinghouse facilities or storage rooms. According
277to previous literature, not only the cultivar, but also the degreening treatment conditions
278and the type of pathosystem clearly affect the influence of degreening on citrus fruit
279susceptibility to postharvest disease. Thus, a decrease in fruit susceptibility to GM was

280observed when oranges were degreened with 5 $\mu\text{L L}^{-1}$ of ethylene at 30 °C and 90-96% RH
281for 3 d (Brown, 1973). In contrast, these degreening conditions significantly increased the
282fruit susceptibility of ‘Robinson’ tangerines to anthracnose caused by *C. gloeosporioides*
283(Brown, 1975). In work by El-Kazzaz et al. (1983a) with oranges, the application of
284exogenous ethylene before fungal inoculation significantly decreased BM severity on
285‘Valencia’ oranges. These authors suggested that longer degreening times reduced
286glucosamine content, and consequently more resistance to the disease, considering that
287glucosamine is an indicator of fungal growth. Likewise, these results showed no polyphenol
288oxidase (PPO) activity in both inoculated and non-inoculated fruit, but phenylalanine
289ammonia-lyase (PAL) activity increased significantly only in the peel of control fruit
290exposed to 1000 $\mu\text{L L}^{-1}$ of ethylene for 6 d.

291 Results from this research work showed that the influence of commercial
292degreening on the development of GM and BM on previously inoculated fruit was
293dependent not only on the cultivar but also on the initial fruit peel CI and the storage
294conditions in which the fruit were held after the 72-h degreening period. Furthermore, the
295effect on the incidence of the molds (% of infected fruit) was generally different than that
296on the severity of the molds (lesion diameter). In general, no significant effect on the
297incidence of both molds was noticed on mandarins and oranges with different initial CI
298artificially inoculated with *P. digitatum* or *P. italicum* before degreening and incubated at
29920 °C for 7 d after degreening. These findings are similar to those obtained by Plaza et al.
300(2004), who observed that degreening with 5-10 $\mu\text{L L}^{-1}$ of ethylene at 20 °C and 90-95%
301RH had no effect on the incidence of BM or GM on ‘Clemenules’ mandarins inoculated
302before degreening. Also, degreening with ethylene gas (5 $\mu\text{L L}^{-1}$ of ethylene for 2 d) did not

303significantly affect the incidence of GM on ‘Valencia’ oranges (McCornack, 1971). Since
304the variable disease incidence measures the amount of infections that take place from free
305conidia deposited in the infection courts (rind wounds) during the artificial inoculation
306procedure, it is clear that ethylene exposure did not affect the germination ability of these
307spores, which remained very high during incubation at 20 °C (Figs. 1, 3, 5). This is in
308agreement with previous results in which the presence or absence of ethylene had no effect
309on the pathogenicity or virulence of the fungus on green-mold-infected citrus fruit (Chalutz,
3101979; Mullins et al., 2000). However, in some of our trials the effect of commercial
311degreening was different on fruit stored at 5 °C for 14 d. Particularly, the treatment favored
312GM incidence on ‘Clemenules’ mandarins with an initial CI of -3.6 and ‘Navelina’ oranges
313with initial CI of -5.3 and 1.7, while it did not affect the incidence of GM or BM in the rest
314of cases. Hence, it seems that in these particular cases, the interaction between previous
315ethylene exposure and prolonged storage at low temperature somehow affected the
316susceptibility of the fruit host allowing a higher amount of spores to germinate and cause
317viable infections. In any case, none of our trials showed a noticeable induction of fruit
318resistance to disease following commercial ethylene degreening of previously inoculated
319fruit. These results differ from those by Porat et al. (1999), who found a 10% decrease in
320the incidence of molds caused by *P. digitatum* and *P. italicum* in response to the application
321of 10 $\mu\text{L L}^{-1}$ ethylene for 60 h on ‘Shamouti’ oranges naturally infected and stored for 4
322weeks at 20 °C. Further, the incidence of GM was reduced by degreening oranges (5 $\mu\text{L L}^{-1}$
323of ethylene at 30 °C and 90-96% RH) for 3 d (Brown, 1973). This dissimilar behavior can
324be logically attributed to important differences in the degreening treatment conditions,
325specifically to different amounts of ethylene and length and temperature of exposure. In

326fact, some research established an involvement of ethylene perception in promoting defense
327responses in citrus fruit infected by *P. digitatum* through the accumulation of defense-
328related mRNAs (Marcos et al., 2005) and global results of transcriptomic analysis of citrus
329fruit peel tissue reveal fundamental effects of phenylpropanoids and ethylene on induced
330resistance (Ballester et al., 2011).

331 Regarding the effect of commercial degreening on disease severity, it was observed
332in these experiments that it highly depended on the cultivar and particularly on initial peel
333CI, while the effect of storage conditions (incubation at 20 °C or storage at 5 °C) was less
334relevant (Figs. 2, 4, 6). The variable disease severity quantifies the growth rate of the
335pathogen once the infection has been initiated. Hence, only the fruit with values of disease
336incidence different than zero were considered for its calculation. Therefore, the effect of
337ethylene degreening on this variable reflects the effect of the treatment not on free spores
338and their capacity to germinate and initiate infection in rind wounds, but on the ability of
339fungal hyphae to grow and multiply in the infection court. The fact that this mycelial
340growth chiefly depends on the fruit host natural resistance to disease, defined basically by
341the genotype and the physical and physiological condition (Palou et al., 2007, 2008), can
342explain the differences in disease severity that were observed in this work. On
343‘Clemenpons’ clementines with different initial peel CI, degreening treatment did not affect
344the severity of the molds, but it significantly increased that of fruit with higher initial CI of
3450.9 for BM on fruit stored at 5 °C (Fig. 2). On ‘Clemenules’ mandarins with lower initial
346CI of -6.5, ethylene degreening did not affect the severity of both GM and BM, while it
347significantly increased that of fruit with higher initial CI of -3.6 or 2.2 (Fig. 4). On
348‘Navelina’ oranges with initial CI of -5.3, degreening exposure did not affect mold severity,
349but the treatment significantly increased the severity of the molds on fruit with higher initial

350CI of 1.1 and 1.7 (Fig. 6). It may be therefore concluded from these observations, the
351results obtained in the previous susceptibility trials performed with fruit with low CI, and
352from the general lack of influence on the variable disease incidence that the effect of
353commercial degreening with ethylene on the development of penicillium molds relied more
354on the natural resistance of each cultivar and on particular fruit condition than on a primary
355influence of the gas on the pathogenic fungus itself. While the peel condition of the
356greenest mandarins and oranges (more immature, lower initial peel CI) was not affected or
357positively affected by commercial degreening in terms of susceptibility to decay, exposure
358to exogenous ethylene of less green citrus fruit (more mature, higher initial CI) significantly
359increased peel susceptibility to disease, presumably through an acceleration of the
360degradation of peel constitutive compounds associated with natural resistance (Porat,
3612008). It is known that the content of major compounds with antifungal activity naturally
362present in the peel of citrus fruit, mostly phenolic compounds such as flavanones, flavones,
363etc. that act as the first line of defense against pathogens, is higher in immature fruit and
364decreases as the fruit ripen and age to senescence, either while remaining in the tree or after
365harvest (Ortuño et al., 2006; Palou et al., 2007). Some postharvest treatments of different
366nature have shown the ability to maintain for longer the effective levels of these
367constitutive antifungals, typically by an enhancement of the activity of defense-related
368enzymes such as PAL, β -1,3-glucanase (GLU), peroxidase (POD), or PPO (Ben-Yehoshua
369et al., 1992; Lu et al., 2013). Since the application of exogenous ethylene can alter or
370accelerate the natural processes of fruit development, ripening and senescence (Kader,
3711985), it could be possible that ethylene degreening contributed to a significant loss of
372bioactive compounds in the rind of more mature fruit (El-Kazzaz et al., 1983b; Sdiri et al.,
3732012b). The influence of the genotype, fruit condition and treatment conditions on the

374effect of ethylene exposure on disease severity can further be explained by contradictory
375results in the literature. In contrast to our findings, work reported by El-Kazzaz et al.
376(1983b), showed that exposure to ethylene at concentrations of 1, 10, 100 or 1000 $\mu\text{L L}^{-1}$
377for 6 d did not affect lesion diameters on ‘Navel’ oranges inoculated with *P. italicum* and
378incubated at 20 °C for 6 d. These authors, however, did not characterize the initial maturity
379of the oranges used for the trials. According to our results, further research should focus on
380the relations between decay severity and initial CI on fruit from each cultivar inoculated
381after degreening. Moreover, future studies might consider the determination of threshold
382values of initial CI for particular cultivars such as ‘Clemenules’ mandarins or ‘Navelina’
383oranges for the significant shift of peel natural resistance to molds under Spanish
384degreening conditions.

385 In this work, ethylene degreening treatment had, in general, no practical effect on
386external quality attributes of citrus fruit, which is in agreement with results reported in
387other research works (Porat et al., 1999; Sdiri et al., 2012b). One exception was the fruit
388firmness of ‘Clemenules’ mandarins with an initial peel CI of -6.5, which was slightly
389increased by ethylene degreening. This result coincided with the findings reported by Plaza
390et al. (2004) on the same cultivar. Likewise, we observed that ethylene degreening had no
391significant effect on internal quality attributes of ‘Clemenpons’ and ‘Clemenules’ mandarin
392fruit. These findings are similar to those obtained on fruits of these cultivars degreened at 2
393 $\mu\text{L L}^{-1}$ for 5 d followed by a cold-quarantine at 1 °C for 16 d (Sdiri et al., 2012b) or on
394‘Satsuma’ mandarin degreened at 4 $\mu\text{L L}^{-1}$ for 5 d (Tietel et al., 2010). In addition, a variety
395of results obtained with different citrus species and cultivar also support our findings (Porat

396et al., 1999; Plaza et al., 2004; Mayuoni et al., 2011a). As one exception, ethylene
397degreening decreased acidity levels in ‘Mosambi’ oranges (Ladaniya and Singh, 2001).

398 We concluded that the effect of commercial degreening with exogenous ethylene on
399the development of citrus penicillium molds relied more on fruit condition than on a direct
400effect of ethylene on the growth of the pathogenic fungi. While the peel condition of the
401greenest (more immature) mandarins was not affected by commercial degreening, exposure
402to exogenous ethylene of less green (more mature) fruit significantly increased peel disease
403severity in some cultivars, presumably through a senescence effect on the fruit peel.

404

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411

412**5. REFERENCES**

413Ballester, A.R., Lafuente, M.T., Forment, J., Gadea, J., De Vos, R.C.H., Bovy, A.G.
414 González-Candelas, L., 2011. Transcriptomic profiling of citrus fruit peel tissue
415 reveals fundamental effects of phenylpropanoids and ethylene on induced resistance.
416 Mol. Plant Pathol. 12, 879-897.

417Ben-Yehoshua, S., Rodov, V., Kim, J.J., Carmeli, S., 1992. Preformed and induced
418 antifungal materials of citrus fruits in relation to the enhancement of decay resistance
419 by heat and ultraviolet treatments. J. Agric. Food Chem. 40, 1217-1221.

420Barmore, C.R., Brown, G.E., 1985. Influence of ethylene on increased susceptibility of
421 orange to *Diplodia natalensis*. Plant Dis. 69, 228-230.

422Brown, G.E., 1975. Anthracnose, a serious decay of degreened 'Robinson' tangerines.
423 Proc. Fla. State Hortic. Soc. 88, 308-311.

424Brown, G.E., 1973. Development of green mold in degreened oranges. Phytopathology 63,
425 1104-1107.

426Brown, G.E., 1986. Diplodia stem-end rot, a decay of citrus fruit increased by ethylene
427 degreening treatment and its control. Proc. Fla. State Hortic. Soc. 99, 105-108.

428Chalutz, E., 1979. No role for ethylene in the pathogenicity of *Penicillium digitatum*.
429 Physiol. Plant Pathol. 14, 259-262.

430Eckert, J.W., Eaks, I.L., 1989. Postharvest disorders and diseases of citrus fruits, In: Reuter,
431 W., Calavan, E.C., Carman, G.E. (Eds.), The citrus Industry, vol. 5. University of
432 California Press, Berkeley, CA, USA, pp. 179-260.

433El-Kazzaz, M.K., Sommer, N.F., Kader, A.A., 1983a. Ethylene effects on *in vitro* and *in*
434 *vivo* growth of certain postharvest fruit-infecting fungi. Phytopathology 73, 998-
435 1001.

436El-Kazzaz, M.K., Chordas, A., Kader, A.A., 1983b. Physiological and compositional
437 changes in orange fruit in relation to modification of their susceptibility to
438 *Penicillium italicum* by ethylene treatments. J. Amer. Soc. Hortic. Sci. 108, 618-621.

439Jiménez-Cuesta, M., Cuquerella, J., Martínez-Jávega, J.M., 1981. Determination of a color
440 index for citrus fruit degreening. Proc. Int. Soc. Citriculture 2, 750-753.

441Kader, A.A., 1985. Ethylene-induced senescence and physiological disorders in harvested
442 horticultural crops. HortScience 20, 54-57.

443Ladaniya, M.S., Singh, S., 2001. Use of ethylene gas for degreening of sweet orange
444 (*Citrus sinensis* Osbeck) cv. 'Mosambi'. J. Sci. Ind. Res. 60, 662-667.

445Lu, L., Lu, H., Wu, C., Fang, W., Yu, C., Yu, C., Ye, C., Shi, Y., Yu, T., Zheng, X., 2013.
446 *Rhodosporidium paludigenum* induces resistance and defense-related responses
447 against *Penicillium digitatum* in citrus fruit. Postharvest Biol. Technol. 85, 196-202.

448Marcos, J.F., González-Candelas, L., Zacarías, L., 2005. Involvement of ethylene
449 biosynthesis and perception in the susceptibility of citrus fruits to *Penicillium*
450 *digitatum* infection and the accumulation of defence-related mRNAs. J. Exp. Bot. 56,
451 2183-2193.

452Mayuoni, L., Tietel, Z., Patil, B.S., Porat, R., 2011a. Does ethylene degreening affect
453 internal quality of citrus fruit? Postharvest Biol. Technol. 62, 50-58.

454Mayuoni, L., Sharabi-Schwager, M., Feldmesser, E., Porat, R., 2011b. Effects of ethylene
455 degreening on the transcriptome of mandarin flesh. Postharvest Biol. Technol. 60, 75-
456 82.

457McCornack, A.A., 1971. Effect of ethylene degreening on decay of Florida citrus fruit.
458 Proc. Fla. State Hortic. Soc. 84, 270-272.

459Moscoso-Ramírez, P.A., Palou, L., 2013. Effect of ethylene degreening on the development
460 of citrus postharvest green and blue molds. Acta Hort. 1012: 633-638.

461Mullins, E.D., McCollum, T.G., McDonald, R.E., 2000. Consequences on ethylene
462 metabolism of inactivating the ethylene receptor sites in diseased non-climacteric
463 fruit. Postharvest Biol. Technol. 19, 155-164.

464Ortuño, A., Báidez, A., Gómez, P., Arcas, M.C., Porras, I., García Lidón, A., Del Río, J.A.,
465 2006. *Citrus paradisi* and *Citrus sinensis* flavonoids: their influence in the defence
466 mechanism against *Penicillium digitatum*. Food Chem. 98, 351-358.

467Palou, L., Crisosto, C.H., Garner, D., Basinal, L.M., 2003. Effect of continuous exposure to
468 exogenous ethylene during cold storage on postharvest decay development and
469 quality attributes of stone fruits and table grapes. *Postharvest Biol. Technol.* 27, 243-
470 254.

471Palou, L., Marcilla, A., Rojas-Argudo, C., Alonso, M., Jacas, J.A., del Río, M., 2007.
472 Effects of X-ray irradiation and sodium carbonate treatments on postharvest
473 *Penicillium* decay and quality attributes of clementine mandarins. *Postharvest Biol.*
474 *Technol.* 46, 252-261.

475Palou, L., Smilanick, J.L., Droby, S., 2008. Alternatives to conventional fungicides for the
476 control of citrus postharvest green and blue moulds. *Stewart Postharv. Rev.* 2:2, 1-16.

477Palou, L., Smilanick, J.L., Usall, J., Viñas, I., 2001. Control of postharvest blue and green
478 molds of oranges by hot water, sodium carbonate, and sodium bicarbonate. *Plant Dis.*
479 85, 371-376.

480Peng, G., Xie, X.L., Jiang, Q., Song, S. Xu, C.J., 2013. Chlorophyll a/b binding protein
481 plays a key role in natural and ethylene-induced degreening of Ponkan (*Citrus*
482 *reticulata* Blanco). *Scientia Hortic.* 160, 37-43.

483Plaza, P., Sanbruno, A., Usall, J., Lamarca, N., Torres, R., Pons, J. Viñas, I., 2004.
484 Integration of curing treatments with degreening to control the main postharvest
485 diseases of clementine mandarins. *Postharvest Biol. Technol.* 34, 29-37.

486Porat, R., 2008. Degreening of citrus fruit. *Tree Forest Sci. Biotechnol.* 2, 71-76.

487Porat, W.B., Cohen, L., Daus, A., Goren, R., Droby, S., 1999. Effects of ethylene and 1-
488 methylcyclopropene on the postharvest qualities of ‘Shamouti’ oranges. *Postharvest*
489 *Biol. Technol.* 15, 155-163.

490Saltveit, J.L., 1999. Effect of ethylene on quality of fresh fruits and vegetables. Postharvest
491 Biol. Technol. 15, 279-292.

492Sdiri, S., Navarro, P., Monterde, A., Benabda, J., Salvador, A., 2012a. New degreening
493 treatments to improve the quality of citrus combining different periods with or
494 without ethylene exposure. Postharvest Biol. Technol. 63, 25-32.

495Sdiri, S., Navarro, P., Monterde, A., Benabda, J., Salvador, A., 2012b. Effect of postharvest
496 degreening followed by a cold-quarantine treatment on vitamin C, phenolic
497 compounds and antioxidant activity of early-season citrus fruit. Postharvest Biol.
498 Technol. 65, 13-21.

499Tietel, Z., Weiss, B., Lewinshon, E., Fallik, E., Porat, R., 2010. Improving taste and peel
500 color of early-season Satsuma mandarins by combining high-temperature
501 conditioning and degreening treatments. Postharvest Biol. Technol. 57, 1-5.

502Zhang, J., 2004. Effect of ethylene on natural resistance of citrus fruit to stem-end rot
503 caused by *Diplodia natalensis* and its relation to harvest control of this decay. Proc.
504 Fla. State Hortic. Soc. 117, 364-367.

505

506 Table 1. Effect of commercial degreening on external quality attributes of citrus fruit

Cultivar	Initial CI	Firmness (% deformation)			Peel break resistance (kgf)			Peel oil release pressure (kgf)		
		At harvest	Control	Degreened	At harvest	Control	Degreened	At harvest	Control	Degreened
'Clemenpons'	-0.07	6.27	8.71a	8.85a	1.23	1.13a	1.09a	-	-	-
	0.9	5.17	5.89a	5.37a	1.35	1.43a	1.27a	4.22	4.93a	3.99b
'Clemenules'	-6.5	4.26	6.02a	5.48b	1.8	1.3a	1.7b	-	-	-
	-3.6	4.42	5.91a	5.87a	1.56	1.46a	1.40a	5.15	5.30a	5.20a
	-0.6	3.44	4.76a	4.53a	1.35	1.57a	1.34a	4.60	4.60a	4.90a
	2.2	3.28	4.56a	4.10a	1.49	1.46a	1.40a	3.75	4.80a	4.46a
'Navelina'	-5.3	1.09	1.51a	1.49a	2.72	2.36a	1.93b	5.90	5.87a	5.40b
	1.1	1.63	2.63a	2.40a	2.11	2.06a	1.80a	5.18	5.76a	5.53a
	1.7	1.60	2.40a	2.03a	2.33	1.86a	1.83a	5.64	5.23a	5.86a
'Nova'	12.3	1.62	2.20a	2.26a	2.17	1.62a	1.70a	5.09	5.75a	5.73a

507 Firmness and peel oil release pressure were measured on 20 fruit per treatment; peel break resistance was
508 measured on 30 fruit per treatment.

509 CI = Color index (CI = 1000.a/L.b; Hunter parameters).

510 -, not determined.

511 For each cultivar and initial CI, mean values within rows followed by the different letters are significantly
512 different according to Fisher's protected LSD test ($P = 0.05$) applied after an ANOVA.

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525 Table 2. Effect of commercial degreening on internal quality attributes of clementine
 526 mandarin fruit

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Treatments	TA (% citric acid)	SSC (%)	Maturity index (MI) ^a	Juice yield (%)
<u>‘Clemenpons’</u>				
At harvest	0.84	11.03	13.21	50.65
Control	1.31a	11.02a	8.41a	52.77a
Degreened	1.36a	11.43a	8.39a	52.24a
<u>‘Clemenules’</u>				
At harvest	0.86	10.53	12.26	45.87
Control	1.31a	10.15a	7.75a	46.80a
Degreened	1.27a	10.47a	8.26a	48.96a

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^a MI = SSC/TA (SSC = soluble solids concentration; TA = titratable acidity).

For each cultivar, columns followed with the different letters are significantly different according to Fisher’s protected LSD test ($P = 0.05$) applied after an ANOVA.

544

545**Fig. 1.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^\circ\text{C}$ and 95-100% RH for
54672 h) on the incidence of green (GM) and blue (BM) molds on ‘Clemenpons’ mandarins
547harvested with different color index (CI = 1000.a/L.b; Hunter parameters), artificially
548inoculated 2 h before degreening, and stored at either $20 \text{ }^\circ\text{C}$ and 90% RH for 7 d or $5 \text{ }^\circ\text{C}$
549and 90% RH for 14 d. For each disease, storage temperature and CI, columns with the same
550letters are not significantly different according to Fisher’s protected LSD test ($P = 0.05$)
551applied to arcsine-transformed data. Non-transformed means are shown.

552

553**Fig. 2.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^\circ\text{C}$ and 95-100% RH for
55472 h) on the severity of green (GM) and blue (BM) molds on ‘Clemenpons’ mandarins
555harvested with different color index (CI = 1000.a/L.b; Hunter parameters), artificially
556inoculated 2 h before degreening, and stored at either $20 \text{ }^\circ\text{C}$ and 90% RH for 7 d or $5 \text{ }^\circ\text{C}$
557and 90% RH for 14 d. For each disease, storage temperature and CI, columns with the same
558letters are not significantly different according to Fisher’s protected LSD test ($P = 0.05$).

559

560**Fig. 3.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^\circ\text{C}$ and 95-100% RH for 4
561d) on the incidence of green (GM) and blue (BM) molds on ‘Clemenules’ mandarins
562harvested with different color index (CI = 1000.a/L.b; Hunter parameters), artificially
563inoculated 2 h before degreening, and stored at either $20 \text{ }^\circ\text{C}$ and 90% RH for 7 d or $5 \text{ }^\circ\text{C}$
564and 90% RH for 14 d. For each disease, storage temperature and CI, columns with the same
565letters are not significantly different according to Fisher’s protected LSD test ($P = 0.05$)
566applied to arcsine-transformed data. Non-transformed means are shown.

567

568**Fig. 4.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^{\circ}\text{C}$ and 95-100% RH for 4
569d) on the severity of green (GM) and blue (BM) molds on ‘Clemenules’ mandarins
570harvested with different color index (CI = 1000.a/L.b; Hunter parameters), artificially
571inoculated 2 h before degreening, and stored at either $20 \text{ }^{\circ}\text{C}$ and 90% RH for 7 d or $5 \text{ }^{\circ}\text{C}$
572and 90% RH for 14 d. For each disease, storage temperature and CI, columns with the same
573letters are not significantly different according to Fisher’s protected LSD test ($P = 0.05$).

574

575**Fig. 5.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^{\circ}\text{C}$ and 95-100% RH for 3
576d) on the incidence of green (GM) and blue (BM) molds on ‘Navelina’ oranges harvested
577with different color index (CI = 1000.a/L.b; Hunter parameters), artificially inoculated 2 h
578before degreening, and stored at either $20 \text{ }^{\circ}\text{C}$ and 90% RH for 7 d or $5 \text{ }^{\circ}\text{C}$ and 90% RH for
57914 d. For each disease, storage temperature and CI, columns with the same letters are not
580significantly different according to Fisher’s protected LSD test ($P = 0.05$) applied to
581arcsine-transformed data. Non-transformed means are shown.

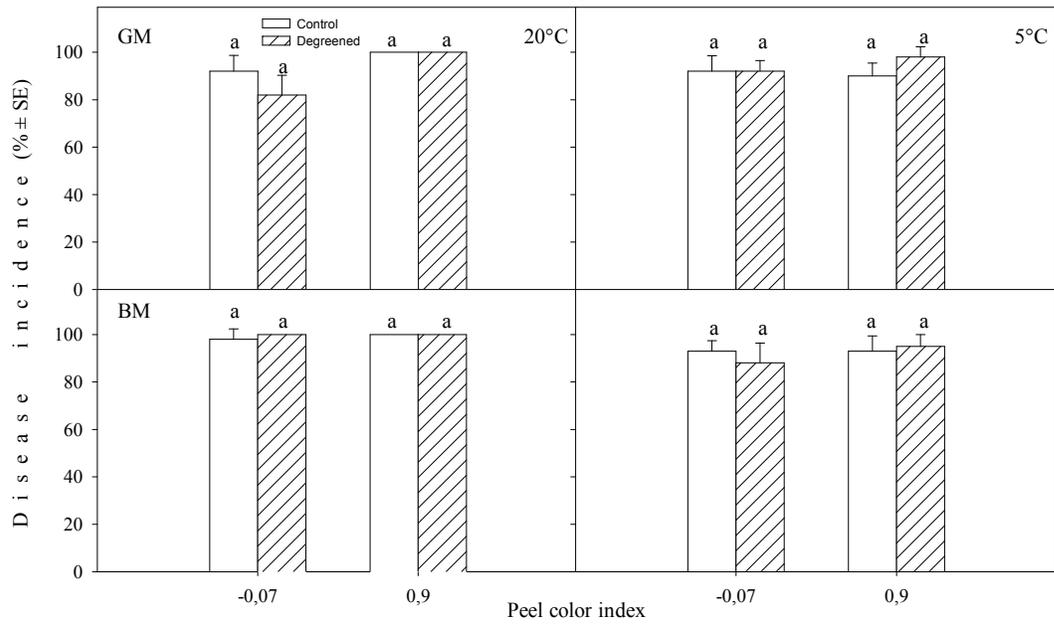
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583**Fig. 6.** Effect of commercial degreening ($2 \mu\text{L L}^{-1}$ ethylene at $21 \text{ }^{\circ}\text{C}$ and 95-100% RH for 3
584d) on the severity of green (GM) and blue (BM) molds on ‘Navelina’ oranges harvested
585with different color index (CI = 1000.a/L.b; Hunter parameters), artificially inoculated 2 h
586before degreening, and stored at either $20 \text{ }^{\circ}\text{C}$ and 90% RH for 7 d or $5 \text{ }^{\circ}\text{C}$ and 90% RH for
58714 d. For each disease, storage temperature and CI, columns with the same letters are not
588significantly different according to Fisher’s protected LSD test ($P = 0.05$).

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592Figure 1.

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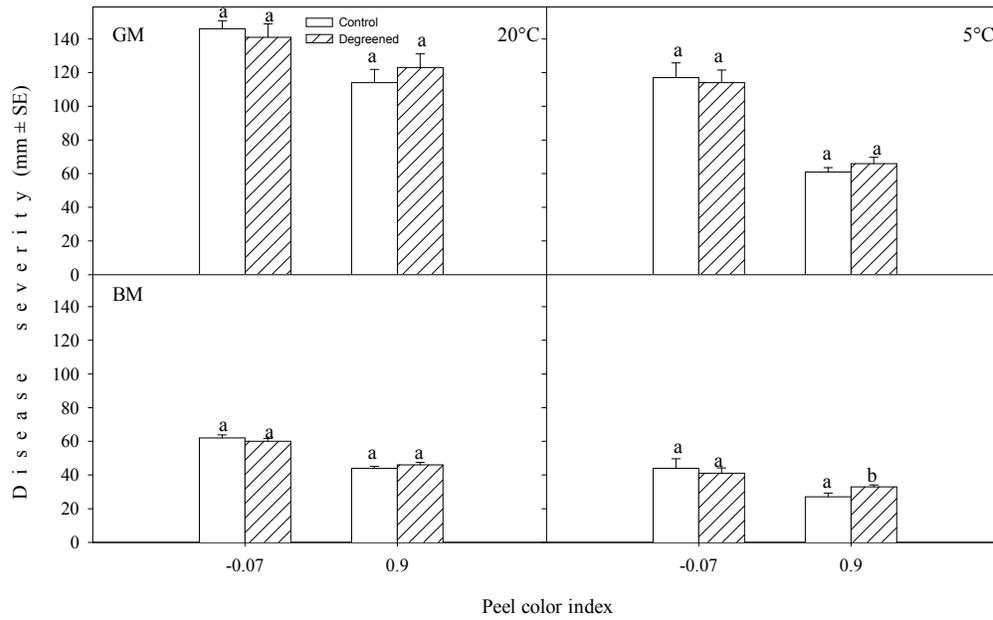
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607Figure 2.

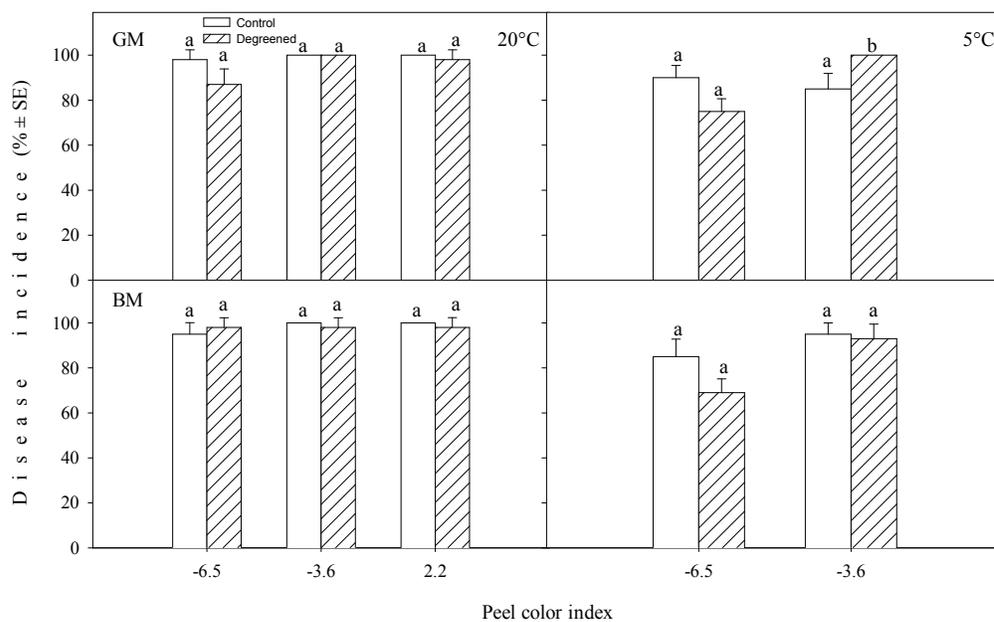
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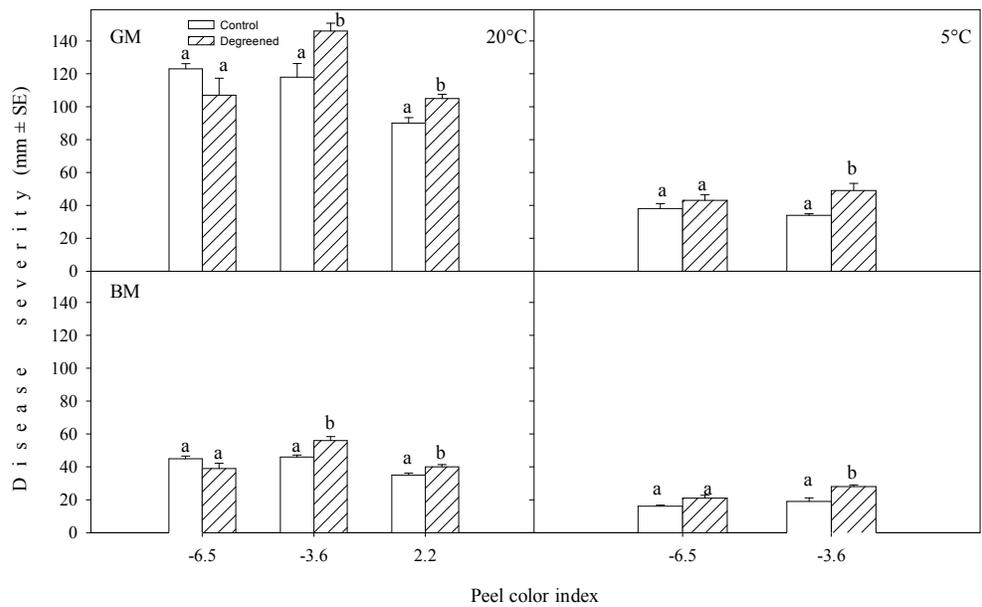


613Figure 3.

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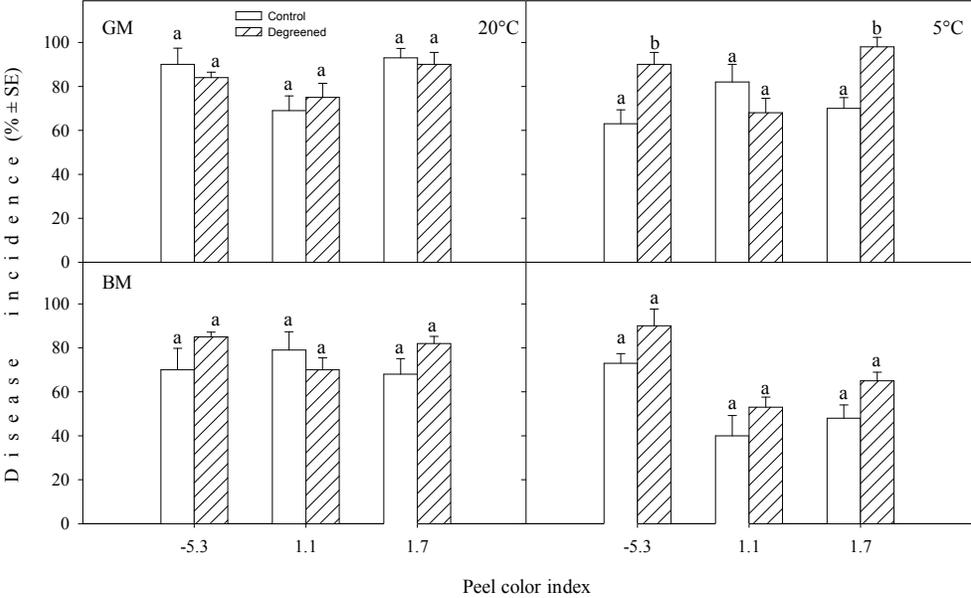


619Figure 4.

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625Figure 5.

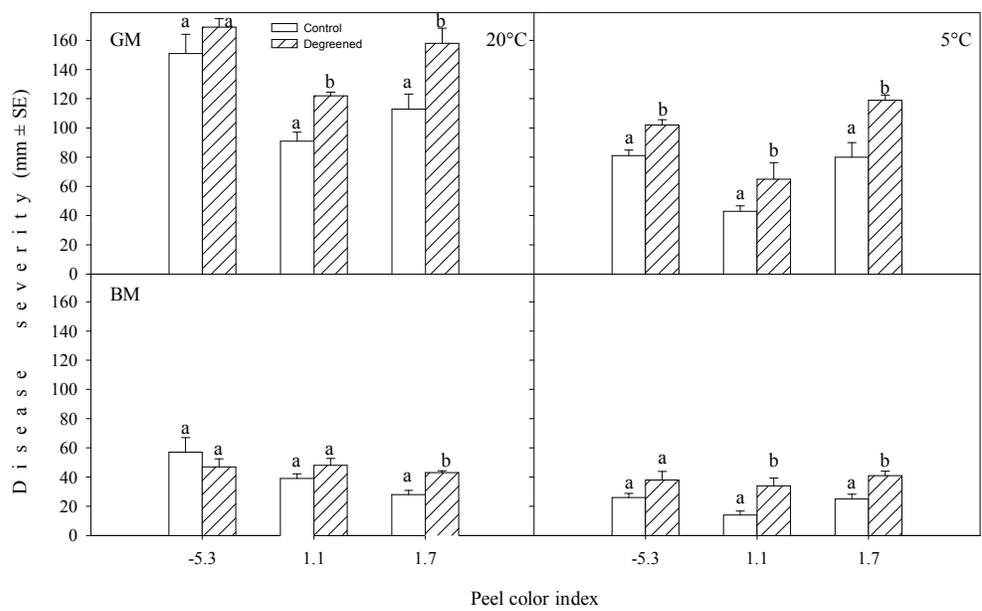
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631Figure 6.

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